

The influence of sleep deprivation on the development of Alzheimer's disease



Illustration by Yen Teoh/Hemera/Thinkstock

Author

Naomi Veeningen

Supervisors

Prof. dr. U.L.M. Eisel & dr. P. Meerlo

June 22nd 2017, University of Groningen

Abstract

Alzheimer's disease (AD) is a neurodegenerative disorder and is the principal cause of dementia in elderly people. The disease is characterized by neuronal senile plaques and neurofibrillary tangles in the patients brain. These plaques consist out of aggregated A β . In 2012 a new clearance system, the glymphatic system, used to remove toxic metabolites has been discovered. The glymphatic system is able to eliminate A β from the brain, hereby preventing it from aggregating. Because the system is highly active during sleep, it is considered that sleep deprivation contributes to an increase in A β . Research is now focusing on the possibility of a relationship between sleep deprivation and the development of AD. This thesis will highlight several possibilities on how sleep, especially sleep deprivation, could influence the development of AD.

Table of contents

1. Introduction.....	4
2. AD Pathology.....	4
3. Mechanisms.....	5
3.1. Amyloid- β hypothesis.....	5
3.2. Tau hypothesis.....	6
3.3. Cholinergic hypothesis.....	7
3.4. Inflammation hypothesis.....	7
4. Sleep and AD.....	8
4.1. Sleep.....	8
4.2. Glymphatic system.....	9
4.3. Aging, sleep deprivation and AD.....	10
4.3.1. Amyloid- β	11
4.3.2. Tau.....	11
4.3.3. Insulin.....	12
4.3.4. ApoE4.....	12
4.3.5. Melatonin.....	12
4.3.6. Orexin.....	13
4.3.7. Immune system.....	13
5. Discussion & Conclusion.....	15
6. References.....	17

1. Introduction

Alzheimer's disease (AD) is a neurodegenerative disorder and is the principal cause of dementia in elderly humans. The disease is characterized by the progressive loss of neurons, which causes the loss of memory and other cognitive functions. Due to the fact there is minimal treatment, AD is a growing public health problem¹⁻³. The economic consequence of AD is immense, the annual cost of dementia in 2010 was valued at 206 billion US dollars^{4,5}. Approximately 70% of the cost arose in Western Europe and North America⁵. At the same time the prevalence of AD was estimated to be 35,6 million cases worldwide. The number of AD cases seems to rise, and is expected to double every 20 years. The amount of AD patients is expected to reach approximately 115,4 million by 2050, unless something changes. The worldwide prevalence for individuals aged 60 or above is estimated between 5 and 7%^{4,6}.

The first case of AD was discovered over 100 years ago by Alois Alzheimer in a patient named Auguste D. Alzheimer described the unusual symptoms of his patient and the presence of neuronal senile plaques and neurofibrillary tangles in the patients brain^{7,8}. A number of breakthroughs have been made since then, for instance the linkage between AD and genetics and the early onset forms of AD⁷. However, only 5% of the AD patients have a clear autosomal dominant (familial) form of AD⁹. While, the remaining 95% occurs with no apparent family history, the so called "Sporadic AD"¹⁰. Sporadic AD does not rise from a genetic disruption, the exact cause is not yet known. The factors involved in this type of AD may be something other than genetics¹¹. For example, disruption of sleep seems to impair the quality of life in AD patients and there appears to be a bidirectional connection between sleep and AD¹². This thesis will review the possible influences of sleep deprivation on the development of AD.

2. AD Pathology

The brain consist out of neurons, the most important cells in the brain. AD causes the loss of neurons, leading to the loss of brain weight and volume³. The pathological processes begin years to decades prior to any symptoms appear^{12,13}. At first soluble amyloid- β ($A\beta$) become insoluble plaques in the brain. These plaques represent the first identifiable pathological change in AD patients, but dementia occurs 10-15 years afterwards¹². The manifestation of the disease starts slowly with mild cognitive impairment. In time, AD will lead to gradual loss of cognitive skills, identity and even activity¹⁰. Some brain regions are more affected then others, mostly affected are the cerebral cortex and the hippocampus¹⁰. As seen in figure 1 the progression of tau and $A\beta$ has a different starting point. Aggregation of tau starts in the locus coeruleus and spreads to the medial temporal lobe, hippocampus and neocortex. The deposition of $A\beta$ starts in the neocortex and spreads to the inside of the brain¹⁴.

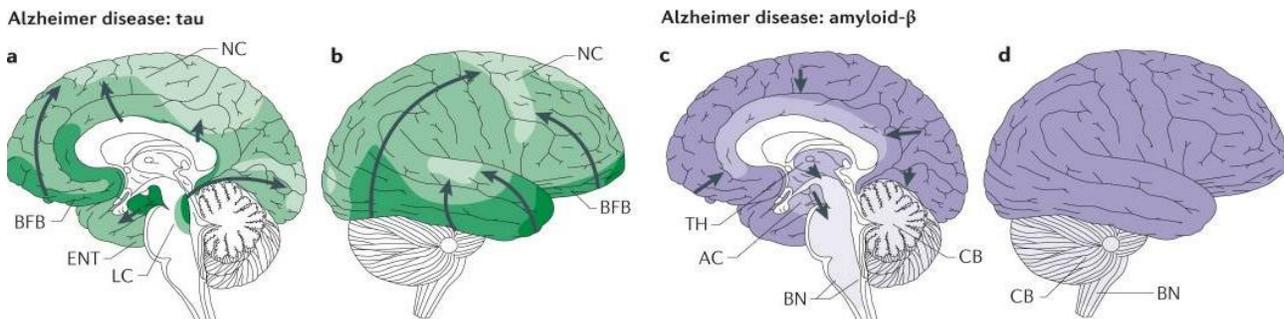


Figure 1 Progression of tau and amyloid-beta in Alzheimer's disease. a,b) Tau aggregation occurs in the locus coeruleus (LC), then in the transentorhinal and entorhinal regions and later in the hippocampus and in the neocortex (NC). c,d) $A\beta$ deposits start in the NC and are then observed in allocortical, diencephalic and basal ganglia structures and in the brainstem¹⁴.

3. Mechanisms

Like multiple other neurodegenerative disorders, AD is associated with the formation and accumulation of abnormally folded proteins. As for AD, there is an increased formation of extracellular amyloid plaques and intracellular neurofibrillary tangles in the brain¹⁵. However, there are different supporting mechanisms of AD pathogenesis and progression. Based on various causative factors multiple theories have been formed, such as the Amyloid- β hypothesis, tau hypothesis and cholinergic hypothesis (fig. 2)^{10,16}.

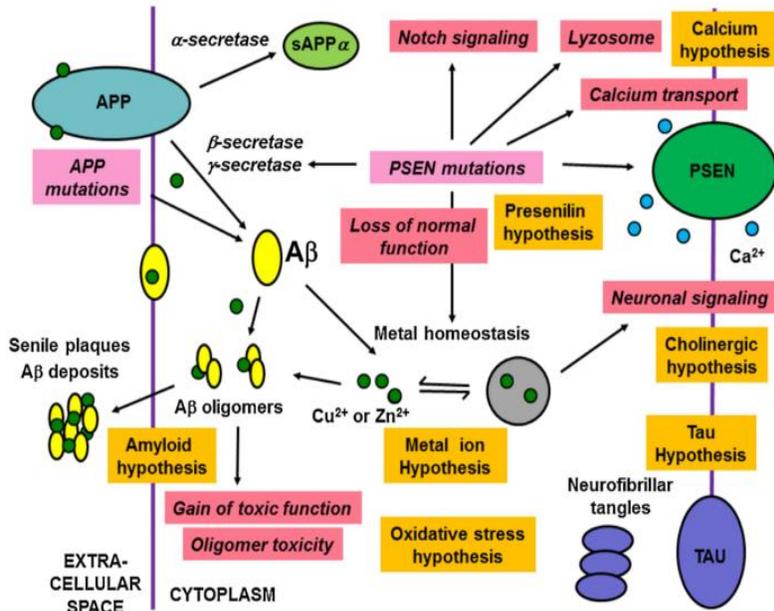


Figure 2 Overview of the biochemistry of Alzheimer's disease. The mechanistic hypotheses are shown in yellow and the key pathologies in red. APP, PSEN and A β are shown in light blue, green and yellow. Metal ions are shown as green or blue

3.1. Amyloid- β hypothesis

The most influential theory for Alzheimer's disease is the amyloid- β hypothesis¹⁷. In which A β is believed to be the main contributor to the dysfunction and loss of neurons^{7,18}. A β is a natural metabolism product and consist of 38 to 43 amino acids^{7,19} and is produced by all cells in the body. Although the main function remains to be determined⁷. Under normal circumstances, the main A β species (A β 1-40) is 40 amino acids long. In case of AD, there is an accumulation of A β 1-42, which is a longer form of A β ⁷.

A β is derived from the amyloid precursor protein (APP) by cleavage (fig. 3)^{7,13,15,19}. Cleavage of APP can occur by two different pathways, the non-amyloidogenic and the amyloidogenic APP processing pathway. The first pathway occurs through cleavage by α -secretase, releasing a large amyloid precursor protein (sAPP α) and leaving an 83-residu carboxy-terminal fragment(C83) behind¹⁹. The cleavage site of α -secretase lies within the A β sequence, α -secretase thus prevents A β formation¹⁸. Afterwards γ -secretase digests C83 and liberates extracellular p3 and the amyloid intracellular domain (AICD)¹⁹. A different pathway is the amyloidogenic APP processing pathway, this pathway is regulated by β - and γ -secretase^{10,19}. One of the main β -secretases is beta-site amyloid precursor protein-cleaving enzyme 1(BACE-1)¹⁸, this enzyme releases a shortened sAPP α . The remaining part, C99, is a γ -secretase substrate, which results in A β and AICD. Then AICD is targeted to the nucleus, where it signals transcription activation¹⁹. The formed A β could be either A β 1-38, A β 1-40 or A β 1-42, depending on the cleavage site of γ -secretase. The degree of aggregation is determent on the type of formed A β , because A β 1-42 displays a higher propensity for aggregation^{10,15}. Researchers also believe that A β 1-42 is a more toxic form compared to A β 1-40, because it aggregates more easily⁷.

A β proteins are able to spontaneously self-aggregate into oligomers (2 to 6 peptides) or fibrils. Fibrils can then be arranged into β -pleated sheets, forming insoluble fibers of advanced amyloid plaques. The soluble forms of oligomers appear to be the most neurotoxic forms or A β ¹⁹. Thus, the severity of the

cognitive defect does not correlate with the total A β amount, but with the levels of oligomers in the brain¹⁹.

The degradation of A β is regulated by proteases. This hypothesis is built on the proposition that AD is caused by an imbalance between A β production and clearance, which results in an increased amount of A β in various forms such as monomer, oligomer, insoluble fibrils and plaques all in the central nervous system²⁰. Proteases such as neprilysin and insulin-degrading enzyme regulate the levels of A β . Neprilysin is known to degrade A β mono- and oligomers. Insulin degrading enzyme is known to degrade small peptides, like insulin and monomeric A β . Overexpression of these enzymes prevent plaque formation^{19,21}.

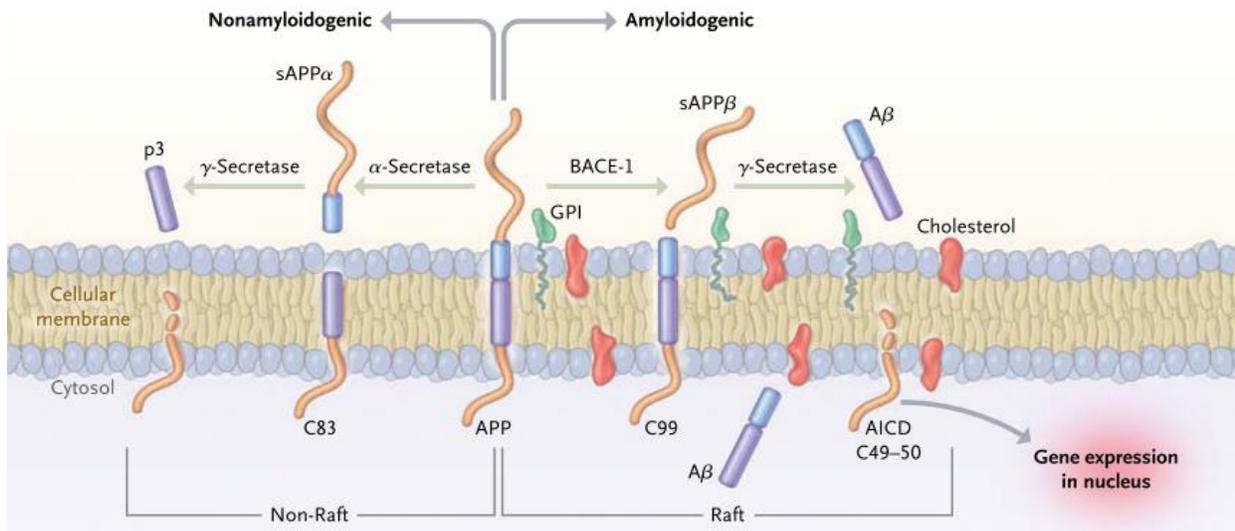


Figure 3 Processing of Amyloid Precursor Protein (APP). Cleavage of APP can occur in two different manners. The non-amyloidogenic pathway (left) occurs through cleavage by α -secretase, generating a large amyloid precursor protein (sAPP α) and an 83-residue carboxy-terminal fragment (C83)¹⁹. The cleavage site of α -secretase lies within the A β sequence and thus prevents A β formation. Following γ -secretase extracts C83 and liberates extracellular p3 and the amyloid intracellular domain (AICD)¹⁹. The amyloidogenic pathway (right) is the second manner through which cleavage can occur, regulated by β - and γ -secretase. The amyloid precursor protein-cleaving enzyme 1 (BACE-1) then releases sAPP β . The remaining C99 is a substrate for γ -secretase, cleavage will generate AICD and A β . AICD will then activate transcription in the nucleus^{10,19}.

3.2. Tau hypothesis

Another important model for AD is the tau hypothesis, since the earlier mentioned intracellular neurofibrillary tangles in the brain are caused by abnormal phosphorylation (hyperphosphorylation) of tau. Tubulin-associated unit (tau) is a microtubule-associated protein (MAP) in neurons, it is an abundant soluble protein¹⁹ and promotes the assembly and stability of microtubules⁸. The gene encoding for tau is microtubule-associated protein tau gene (MAPT)^{3,8}.

The cytoskeleton of the neuron is composed of microtubules, which maintain the neuronal structure, axonal transport and neuronal plasticity²². These microtubules are vital for transport of organelles and vesicles containing proteins and neurotransmitters, which are transported from the neuron cell body (soma) to the synapses²². Microtubules are formed by accumulation of α - and β -tubulin, this accumulation is stabilized by phosphorylated tau^{8,22}. The stabilizing activity of tau is regulated by its degree in phosphorylation⁸. Enzymes regulate the extent of tau phosphorylation¹⁹, increases

phosphorylation by glycogen synthase-3 (GSK3) activity and decrease by phosphatases (PP2A and PP2B)⁸. In case of AD, tau is abnormally hyperphosphorylated²², this is caused by kinases, like GSK3, and affects its ability to bind to tubulin⁸. Resulting in the detachment of tau^{4,8} and thus destabilizing the microtubule structure⁸. Hyperphosphorylated tau is, in contrast to normal tau, insoluble and self-aggregates into paired helical filaments(PHF)¹⁹. Neurofibrillary tangles consist of these PHF's and thus tau¹⁶.

Interestingly, there is evidence that A β oligomers induce oxidative damage and promote tau phosphorylation^{16,17}. What seems to suggest that hyperphosphorylation of tau and the formation of neurofibrillary tangles in the neurons are secondary events¹⁷

Several kinases are implicated in the hyperphosphorylation of tau. Hyperphosphorylation can also occur via the extracellular signal-regulated kinase (ERK) pathway and may occur under conditions where oxygen is lacking, like hypoxia. Whereas hypoxia is a common feature of Obstructive Sleep Apnoe(ASO), one of the most common sleep disorders. This disorder could trigger neuronal degeneration and axonal dysfunction in the cortex and brainstem²³.

3.3. Cholinergic hypothesis

In the late 1960s and early 1970s, the first systematic biochemical investigation of the AD brain began. This is how researchers considered the cholinergic hypothesis. In the mid-1970s researchers reported a neocortical deficit in Choline acetyltransferase (ChAT), which is responsible for the synthesis of acetylcholine(ACh). This deficit was confirmed by discoveries of reduced ACh release, choline uptake and loss of cholinergic soma in the nucleus basalis of Meynert, the cortex and the hippocampus²⁴⁻²⁸. Thereafter, it prompted researchers to believe that the loss of neurons was due to the loss of cholinergic stimulation²⁹.

The neurotransmitter ACh is involved in learning and memory³⁰. ACh release and synthesis are depressed in AD and the ACh degradation is altered in the presence of A β ³¹. Furthermore, attachment of amyloid fibers to Acetylcholine esterase (AChE), catalyzes the breakdown of ACh²⁴⁻²⁸, can trigger an alteration in its characteristics. For example, changing its pH sensitivity and thus its activity³². Apart from this, it has been noticed that the AChE levels in AD patients are decreased, while its activity is increased around plaques and NFT³³.

To test this hypothesis researchers used AChE inhibitors, which could reduce the hydrolysis of ACh. Testing these AChE inhibitors, patient's showed a lot of side effects. To fairly test the cholinergic hypothesis, scientists developed a 'second generation' of AChE inhibitors^{34,35}. The use of these inhibitors showed a delayed progression of the symptoms of AD, equal to 6-12 months deterioration³⁶⁻³⁸. The fact that the AChE inhibitors only reduce the progression of the disease indicates that the cholinergic dysfunction may not cause AD related cognitive impairment directly^{35,39}.

3.4. Inflammation hypothesis

Another hypothesis is the inflammation hypothesis. Neurodegenerative diseases are all accompanied by the activation inflammatory and neuroinflammatory systems. The inflammation hypothesis for AD is built on assumptions that the immune system accompanies the AD pathology and could be a contributor to the pathogenesis of the disease^{40,41}. The link between inflammatory processes and the pathogenesis of AD has been supported by epidemiological studies, describing a reduced risk in the development of AD when non-steroidal anti-inflammatory drugs are used chronically⁴².

Neuroinflammation is characterized by the activation of microglia and astrocytes. Astrocytes are part of the blood-brain barrier, functioning as the modulators of structure and of the brain⁴³⁻⁴⁵. They play an important role in the trophic and metabolic support of neurons, neuronal signal transmission and in the formation and plasticity of the synapses⁴⁶⁻⁴⁹. The microglia are considered the principal immune cells of the central nervous system (CNS), functioning as the macrophages of the brain by the ability to migrate into different parts of the brain, phagocytose, process and present antigens^{45,50-52}. They can be triggered by detection of protein aggregation or neuronal death. In case of AD, the microglia are able to bind to soluble A β oligomers and A β fibers through receptors. This process is followed by phagocytosis.^{53,54} In cases of sporadic Alzheimer, downregulation of expression of A β phagocytosis receptors are suggested to be responsible for inefficient clearance of A β .^{53,55} Normally, the presence of A β can prime microglia, making them susceptible to a secondary stimulus or promoting their activation. In AD, microglial cells are chronically activated by A β ^{56,57}. Resulting in continuous production of inflammatory cytokines and chemokines^{41,58,59}. Whereas, cytokines and chemokines maintain the activation of primed microglia. This process affects surrounding CNS cells: astrocytes, oligodendrocytes and neurons. Possibly leading to the hyper phosphorylation of tau, and thus leading to neurodegeneration^{41,45}. Adding to this, that it is possible that these inflammatory processes contribute to the sleep associated development of neurocognitive disorders⁶⁰

4. Sleep and AD

It has been noticed that sleep of patients with AD is altered^{12,61}. It is even suggested that fragmented sleep could be an early sign of Alzheimer or even a contributor to the development of the disease. This chapter will explain if the bidirectional relationship is possible and the research that does and doesn't support it. However, first the processes underlining sleep will be described.

4.1. Sleep

Sleep is a fundamental biological process, commonly observed throughout the animal kingdom. Although, the systems generating sleep are poorly understood. Sleep takes up to one third of our lifetime, which could mean it plays a special role in our existence, otherwise it would have disappeared through evolution. Insufficient sleep can impair our cognitive performance during wakefulness⁶². Long term or chronic sleep deprivation is linked to many health issues, for example obesity, cardiovascular diseases and neurocognitive disorders^{4,62}. The exact mechanism or order of incidence is not always known.

Sleep consists of two types of sleep, rapid eye movement (REM) and non-REM⁶². NREM consists of three stages (N1, N2 and N3), which are progressively deeper. Stage one is a transition state consisting of light sleep, at this point there is a reduced brain-wave activity and slow eye movements. In stage two the body is in a low energy state; muscle relaxation, decreased body temperature and a reduced heart rate. This stage is defined by spindles and K-complexes, less than 3 min apart on electroencephalographs. In deep sleep, stage 3, sleep is characterized by slow-wave-spindles (SWS). This stage of sleep is thought to be the most restorative form of sleep. The next stage, REM, is characterized by Rapid Eye Movement, vivid dreaming, active inhibition of muscles and an increased brain activity, heart rate and respiratory rate. Overall, adults go through all four stages in 90-100 minutes during the night. However, the time spent in one stage changes as the night progresses^{4,63-65}.

4.2. Glymphatic system

Until a few years ago $A\beta$ was thought to be cleared only by the following mechanisms; uptake by microglial phagocytes, receptor-mediated transport across the blood vessel walls and degradation by enzymes⁶⁶. Comparable mechanisms occur for tau⁶⁷. However, in 2012 the discovery of the glymphatic system provided new insights in the clearance of the brain⁶⁸⁻⁷⁰.

The glymphatic system is a biomolecule clearance system used to remove toxic metabolites in the brain via the cerebrospinal fluid (CSF) and interstitial fluid (ISF)⁷¹. The CSF is found in the brain and spinal cord and is produced by the choroid plexuses of the ventricles of the brain. The interstitial fluid is located between tissue cells⁷⁰⁻⁷². However, the glymphatic system does not just eliminate soluble proteins and metabolic waste. The system also takes care of distribution, e.g. lipid distribution by releasing Apolipoprotein E⁷³. Though this is literature study will focus on the clearance of biomolecules, since this is a potential important mechanism for AD.

The glymphatic system clears biomolecules from the brain by a convective flow (fig 4). The CSF enters the brain parenchyma via a para-arterial influx. The movement of CSF into the parenchyma drives

interstitial fluid between tissue towards the perivenous spaces^{72,74,75}. This way the extracellular solutes are removed through drainage of the interstitial fluid along the para-venous efflux to the cervical lymph system via the olfactory bulb and along cranial and spinal nerves. The interchange between CSF and ISF is driven by a combination of respiration, arterial pulsatility and CSF pressure occurring through constant production of CSF by the choroid plexus^{69,70,72}. The transport of CSF into the brain parenchyma is facilitated by Aquaporin 4 (AQP4) water channels⁶⁹⁻⁷². The AQP4 water channels are expressed in the endfeet of astrocytes located in the brain vasculature^{69,70,72}. The channels are required for glymphatic functioning and facilitate clearance of soluble proteins, waste products and excess extracellular fluid⁶⁹⁻⁷¹. The glymphatic system takes care of 65% of the $A\beta$ clearance, most of which occurs during sleep. It has been tested in mice that clearance occurs twice as fast in sleeping and anesthetized fase^{69,76}. Natural sleep and anesthesia facilitate a 60% increase in interstitial space, leading to a convective flow between CSF and ISF^{76,77}. The increase in interstitial space leads to an increased clearance of $A\beta$ ^{69,76}.

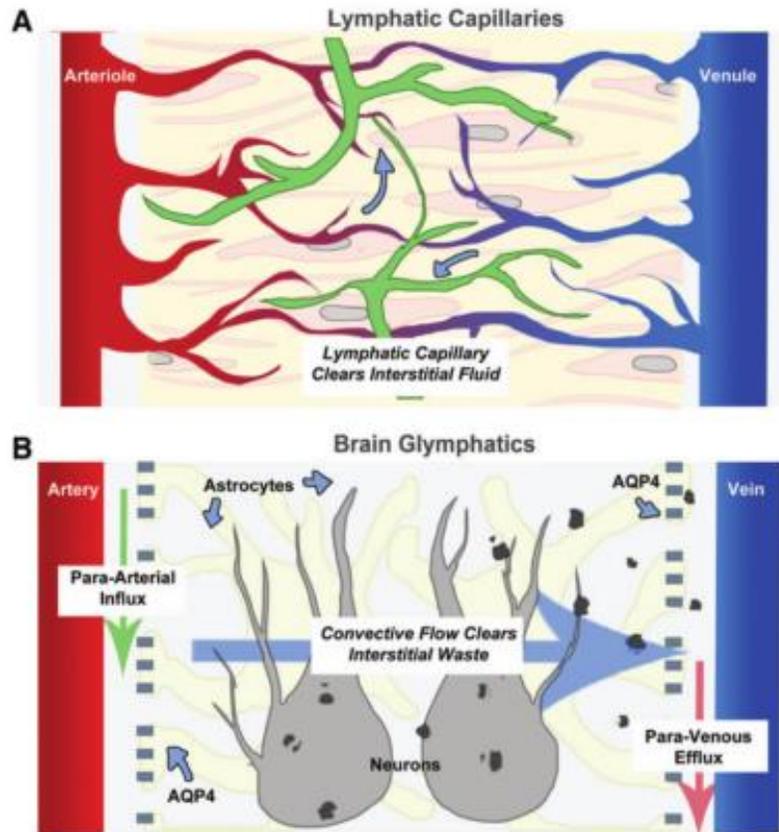


Figure 4 The lymphatic system is comparable to the glymphatic system. The lymphatic vessels eliminates waste from the interstitial fluid(A). In the brain, the glymphatic system, there is a convective flow between entering cerebrospinal fluid through the arteries and exiting interstitial fluid through the veins removing biomolecular waste from the brain⁷¹.

The biomolecular clearance is probably not regulated by circadian rhythms, due to the fact that anesthesia can be administered at any time inducing clearance. One hypothesis is that arousal itself is responsible. To be precise, the locus coeruleus noradrenergic signaling, which is linked to wakefulness. Thus, adrenergic signaling is possibly able to reduce interstitial volume followed by a decreased clearance. After treatment of adrenergic antagonist, mice showed an increase in CSF influx as well as an increase in interstitial space. The researchers even administered more slow wave patterns, indicating a more relaxed sleep-like state⁷⁶.

4.3. Aging, sleep deprivation and AD

As people age, it is often seen that people need more time to fall asleep, wake up more often and earlier and have excessive daytime napping^{4,78,79}. Additionally, Elders tend to have a reduced threshold for arousal resulting in fragmented sleep with multiple arousals⁶⁵. Aging individuals also seem to awake less from REM sleep and more from NREM sleep, compared to young adults, and have less sleep spindles and K-complexes in NREM sleep^{4,78,80,81}. Studies have indicated that aging is associated with reduced SWS and NREM⁸², this occurs in combination with structural brain atrophy (mainly in the frontal lobe regions)⁸³. A study in 2013 even demonstrated that the degree of age-related atrophy in the medial prefrontal gray matter is linked to the degree of reduced SWS activity⁸⁴. Furthermore, Mander et. al hypothesize that A β accumulation within the medial prefrontal cortex (mPFC) is significantly correlated with the severity of impairment slow-wave activity(SWA) during NREM sleep⁸⁵.

As for the link between SD and AD development. It has been reported that excessive daytime sleepiness, at a baseline, have a higher probability to be diagnosed with incident dementia later on, compared to those without EDS. Likewise, individuals with sleep disturbances were more likely to develop or be diagnosed with AD or cognitive decline at a follow up⁸⁶⁻⁸⁸. Another study found that reduced sleep was associated with a 75% increased risk for all cause dementia and doubled the risk for AD. In which no effect of sleeping pills has been administered, because they may not result in quality sleep⁸⁹. Another study also underlined that markedly fragmented sleep could increase the risk at developing AD, by 1,5 fold, compared to those with the smallest amount of fragmented sleep⁹⁰. Other studies in humans also suggested that sleep disruption and disorders could be a possible risk factor for the development in cognitive deterioration and dementia^{88,91}. A study in 70 year old's reported a correlation between shorter sleep duration or poorer sleep quality and greater A β burden assessed by PET⁹².

In AD patients imaging and pathological studies showed abnormalities in brain regions that are known to regulate sleep⁹³. This explains the changed sleep pattern seen in AD patients. Even at early stages, before clinical onset, disruptions of NREM and SWS have been detected^{85,94}. In patients with AD, insomnia is very common, the severity of this symptom even correlates with the degree of dementia⁹⁵. As explained before, sleep facilitates in the clearance of metabolic waste by the glymphatic system⁷⁶. With the changed sleep pattern this clearance mechanism could be suppressed¹². The amount of A β is directly affected by how much people are awake or asleep. Likewise, it has been reported that individuals with AD and sleep disruption are more likely to become symptomatic compared to AD patients with a healthy sleep pattern¹². Correspondingly, animal models for AD have shown that manipulation of sleep and circadian behavior can modify the progression of the disease. As sleep deprived mice had 25% more β -amyloid plaques compared to control mice, implying that disruption in the sleep-wake cycle and orexin may play a role in the pathogenesis of AD⁹⁶.

4.3.1. Amyloid- β

The release of A β is facilitated by neuronal firing, or synaptic activity⁹⁷. A β concentrations show diurnal variation, the levels rise during wakefulness and fall during sleep^{2,98,99} in mice⁹⁹ and humans⁹⁸, due to its glymphatic system. Therefore, it has been suggested that sleep facilitates in the removal of A β ^{4,99,100}. Neurons are predicted to have the less neuronal activity during sleep, especially during SWS and thus release less A β compared to other stages of sleep or wakefulness. In case the quality of sleep is poor and an individual may not reach and sustain SWS, this could result in a greater release of A β ⁹⁹ and possibly to A β accumulation¹⁰⁰.

In case of sleep deprivation, the amount of A β will not decrease as much compared to normal sleep, which may be important in the pathogenesis of AD. A research by Ooms et al. (2014) indicates that one night of sleep deprivation increases cerebral A β 42 levels, which could elevate the risk at AD. Though this research also found that one night sleep deprivation does not increase the levels of A β 40 and tau¹⁰⁰. Furthermore, Di Meo et al. (2014) investigated the impact of sleep deprivation on an AD mouse model with plaques and tangles and found significant memory impairments, altered tau metabolism and synaptic pathology¹.

Transgenic models of AD have shown that environmental or pharmacological manipulation of sleep and circadian behavior can modify disease progression^{96,99,101}. Furthermore, it is seen in mice that the diurnal A β level variation seems to disappear when A β plaques are present and this also applies to humans, particularly the levels of A β 42 remains even⁹⁸. Evidence has shown that individuals with plaque formation have worse sleep quality compared to individuals without plaque formation. The affected subjects, have a lower sleep efficiency and the wake up time after sleep onset is affected¹⁰². However, it is not known whether the plaque formation is a result of reduced sleep quality or the other way around. Though, it has been said that chronic sleep deprivation accelerates A β deposition into insoluble A β plaques⁹⁹.

4.3.2. Tau

Evidence has shown that tau indirectly participates in the regulation of the sleep and wake cycle. Tau deficient mice show an increase in wakefulness and a decrease in NREM sleep time, a higher number of state transitions from NREM to wakefulness and shortened sleep bouts¹⁰³.

Microtubules are able to modulate the sensitivity states of different melatonin receptors, and can thereby influence the activity of circadian patterns^{104,105}. As a result, impaired functioning of melatonin receptor could trigger sleep disturbances in AD patients. This finding is supported by the discovery of increased wakefulness in tau deficient mice¹⁰³.

A research by Holth et. al (2017) also indicated that tau pathology is associated with sleep disturbances. This research showed not only a decrease in NREM sleep but also a decrease in REM sleep. Tau related changes in sleep could negatively affect disease progression. As well as A β , tau levels in the brain interstitial fluid increase by neuronal activity. Suggesting it could be a possibility that increased wakefulness may elevate tau secretion and maybe even worsen tau pathologies¹⁰⁶. Though, the function of tau in the extracellular space is not well understood^{107,108}.

Sleep deprivation has shown to reduce the phosphorylated isoform of soluble tau, which is related to the increase of its insoluble form. These data suggest that SD could accelerate the tau pathology, by inducing conformational changes and thereby accelerating tau aggregation¹. This change could be associated with

the reduced activity of cdk-5, a kinase involved in the post-transcriptional phosphorylation of tau and AD^{109,110}. Considering the contradiction between the cdk5 activity and the percentage of phosphorylated tau, it is believed that other kinases and/or phosphatases may be involved. Though, it is also possible that the lower percentage of phosphorylated tau results from hyperthermia, which is a consequence from SD¹. Another study in a mouse model for AD showed elevated A β and tau-p levels in the cortex (not hippocampus) after chronic mild sleep restriction¹¹¹.

4.3.3. Insulin

Sleep deprivation has not only been associated with a reduced glymphatic metabolism, but also with the glucose metabolism. Research has shown that the glucose metabolism is also altered in sleep deprived patients, contributing to increased blood glucose and an decreased insulin level, which can lead to insulin resistance like type 2 diabetes mellitus¹¹². This insulin resistance, has already been observed after one night of SD in healthy subjects¹¹³. Interestingly is the fact that diabetes mellitus is also associated with an increased risk for the development of AD^{114,115}. Though, it has been shown that insulin resistance is not always involved in the pathogenesis of AD. For instance, a mouse model carrying an insulin receptor mutation did not display acceleration in plaque formation or memory abnormalities¹¹⁶.

4.3.4. ApoE4

ApoE4 is a major risk factor for late onset AD, especially in homozygous cases¹¹⁷ with an increased risk of 50%¹¹⁸. ApoE is thought to play a critical role in A β clearance, through binding to A β . ApoE4 is known to form less stable complexes compared to ApoE3 and ApoE2, hereby decreasing A β clearance.

Some studies propose that ApoE4 could make MCI or demented patients more susceptible for sleep alterations (e.g. reduction of REM sleep, increased fragmentations of SWS or sleep apnea)^{94,119,120}. Decreased sleep efficiency could be due to a lower CSF melatonin rate in homozygous 4/4 compared to heterozygote patients¹²¹. Other studies associate homozygous 4/4 genotypes with an even more increased risk of developing sleep disorders and cognitive impairment related to sleep diseases^{120,122–124}. A study among older women indicated that sleep disordered breathing could be associated with the increased risk of developing dementia, particularly if the allele Apolipoprotein E epsilon4 is present.^{91,125}

Though, other research has shown a protective effect of ApoE4 on sleep in AD¹²⁶, or either no effect⁶¹. However, it has been shown that better sleep consolidation could reduce the risk on AD conferred by ApoE4¹¹. Though, research has indicated that the negative effect of ApoE4 might be amplified by sleep disruption^{92,123,124,127,128}.

4.3.5. Melatonin

Melatonin is a hormone produced by the pineal gland at night. This hormone plays an important role in the regulation of the biological clock and sleep wake cycle, by promoting sleep onset⁴. Melatonin has a protective role as an antioxidant and anti-amyloid properties as well. It can arrest the formation of amyloid fibrils and is able to attenuate the AD-like hyperphosphorylation of tau. Melatonin also seems to play a role in the protection of the cholinergic system and in anti-inflammation^{129,130}.

Research has shown that the total level of melatonin in the body decreases with age¹³¹. In AD patients levels of melatonin are even more decreased¹³², these reduced levels of melatonin correlate with the severity of mental and sleep impairments^{133,134}. Interestingly enough, the homozygous ApoE4 patients display a significantly lower amount of melatonin compared to patients with only one copy¹²¹.

The results supplementation of melatonin to AD patients have been contradictory. Some studies have shown little to no effect on sleep in AD patients^{135,136}. Though, these patients had more severe AD, with probably a higher level of A β . As for normal aged individuals, improving effects have been seen¹³⁷. Though, there are studies that do report a reduced variability of sleep onset time and a reduced percentage of nighttime activity. Especially long-term administration of melatonin showed improvement of sleep quality in AD patients¹³⁸⁻¹⁴⁰. Even the cognitive alterations in AD patients seemed to be halted in melatonin receiving patients compared to non-receiving patients^{139,141}. Studies with APP transgenic mice have indicated that early long-term melatonin treatment provides anti-amyloid and antioxidant effects. However, it is seen that when treatment is given after amyloid formation there is no such effect¹⁴²⁻¹⁴⁴. Melatonin seems to be useful in symptomatic treatment, concerning sleep, sundowning (agitated behavior during the evening) and cognitive impairment¹⁴⁵. As for sleep specific phases, melatonin treatment has shown to improve REM sleep in elders¹⁴⁶ and is found to improve the restorative phases of sleep in AD patients^{145,147}. Which is interesting considering the decrease in REM sleep in AD patients¹⁴⁸.

4.3.6. Orexin

Orexin (hypocretin) is a neurotransmitter that is responsible for regulating arousal, wakefulness and is able to suppress REM sleep. The loss of orexin-producing neurons causes narcolepsy⁹⁹. Furthermore, it has been reported that the orexin system is affected in advanced AD, could be a consequence of the loss of orexin-producing neurons¹⁴⁹. Since the glymphatic system rinses the brain from toxins during sleep, excessive wakefulness regulated by orexin could possibly be a key regulator in AD.

For instance, infusion of a dual orexin receptor antagonist (almorexant) led to a decreased A β level in the interstitial fluid in mice and to a 10% decrease in wakefulness. Research shows that the knockout of the wake-promoting orexin gene reduces A β accumulation. This effect is reversed by sleep deprivation¹⁵⁰. Indicating that sleep might be a modulatory factor for the degree of A β toxicity¹⁵¹. Which, suggest that chronic sleep deprivation may contribute to the pathogenesis of AD⁹⁹. Though, patients whom have narcolepsy-cataplexy (thus, orexin deficient) don't seem to be protected against AD¹⁵², which is interesting considering the effect of orexin antagonist's in AD mice.

4.3.7. Immune system

Sleep deprivation will provide a deregulation of the immune system. Study findings in general, suggest that SD is accompanied by immune activation. For example, increased white blood cells, granulocytes, monocytes, lymphocytes, NK cells and NK activity in the blood circulation have been reported¹⁵³. Studies also found a changed balance in cytokine production. Some research suggested that the production of IL-6 and TNF- α is increased by prolonged sleep restriction^{154,155}, for which saturable transport systems over the blood-brain barrier are described¹⁵⁶. There are also studies that outlined the effects of 24 h of continued wakefulness on cytokine production. Though these result are very conflicting^{157,158}. Some research found increased levels of IL-1, IL-2 and IL-10^{155,159}. Which is interesting, considering the firing rate of wake-active serotonergic neurons are reduced by IL-1 through enhancing inhibition of the axon terminals¹⁶⁰. Furthermore, studies have shown that IL-1 and TNF are able to increase NREM sleep^{161,162}, which promotes immune function and occurs SWS. IL-1 is also found to cause fragmentation of NREM sleep¹⁶³, in which it's effect magnitude and duration depends on the dose and time and time of administration¹⁶³⁻¹⁶⁵. Highlighting the contradictory of these studies.

Research highlighting the role of glial cells and SD revealed some interesting results. For example, a recent study by Bellesi et al. (2017) found that after normal sleep, astrocytes are active in 5,7% of the brain's synapses of mice. While, in sleep deprived mice, astrocytes were active in 8,4% of the synapses compared to 13,5% in chronically sleep deprived mice. Suggesting that chronic sleep loss results in more debris breakdown, but also in an elevated demolition of neuronal connections in the brain. Furthermore, their research found that after chronic sleep deprivation even microglial cells were more active. Excessive activation of microglial cells has also been seen in AD¹⁶⁶. Previously, research had already reported astrocytic activation in the hippocampus of sleep deprived rats¹⁶⁷.

Inhibition of microglial activation in a study by Wisor et al. (2005) resulted in a reduced rebound in SWS after 3 hours of SD. Hereby they suggested that the microglial activation could contribute to the need for sleep⁹⁶. Although another study by Bellesi et al. (2015) presented that 6-8 h of sleep deprivation led to an increased rebound of SWS activity, without finding evidence for microglial activation¹⁶⁸. Which could indicate that microglial activation does not necessarily play a role in sleep homeostasis. The activation, however, could be a physiological response to worn synapses by extended wakefulness. Though, this could also be due to the reduced clearance of A β by the glymphatic system as a result of SD⁷⁶, leading to the aggregation of A β ⁹⁹, by which microglial cells can be activated^{169,170}.

5. Discussion & Conclusion

Looking at the overall findings it is clear that AD is a complex disease, in which multiple factors can play a major role. In all of the AD subjects tangles and A β plaques are important markers for the disease^{7,8,15}. Because the cause of sporadic AD is not yet identified, this literature research focused on the possibility of sleep deprivation as a risk factor for AD.

Sleep is known to facilitate the removal of toxic metabolites, like A β molecules, from the brain using the glymphatic system. Sleep deprivation could reduce the clearance of A β by the glymphatic system^{68-71,76,100}. Furthermore, aging is associated with a reduced sleep quality^{4,65,78,79}, while aging is also associated with an increased risk of AD¹⁹. Elders seem to have a reduced SWS and NREM sleep⁸². As for AD patients, a decrease in SWS and REM sleep and some alterations in spindles and K complexes has been noted¹⁴⁸. Considering that SWS is thought to be the most restorative state of sleep, because there is less activity of the neurons, a decrease in sleep and SWS sleep could result in a higher amount of A β levels in the brain^{4,63-65}. Apart from speculating, research has shown that AD patients with sleep disruptions are more likely to become symptomatic than patients with a healthy sleep pattern¹². Adding to this that sleep deprived animal models for AD have a 25% increase in A β plaques compared to non-sleep deprived controls⁹⁶.

As for tau, it is seen to indirectly regulate the sleep and wake cycle. Tau deficiencies and tau-p appears to increase wakefulness and decreases NREM and REM sleep^{1,103,106}. Because tau is thought to increase neuronal activity, presence of tau in the ISF could negatively affect AD progression¹⁰⁶. Furthermore, SD has shown to result in elevated tau-p levels¹¹¹.

SD has also been associated with reduced insulin level, possibly leading to diabetes mellitus¹¹². Which is thought to increase the risk of AD^{114,115}. Though one research has indicated that an insulin receptor mutation in mice did not exacerbate AD-like phenotypes¹¹⁶.

For people carrying the ApoE4 gene, research has shown that better sleep could reduce the risk of AD¹¹. And that increased fragmented sleep in ApoE4 carriers increase the risk at AD by 1,5 fold⁹⁰. Indicating that sleep is important in the development of AD for people at risk nonetheless.

Levels of melatonin seem to decrease in elders¹³¹ and even more in AD patient¹³². Melatonin supplementation studies have shown contradictory results^{135,136,138-140}. Though, it could be that long term administration showed better improvement. Furthermore, it could be possible that in severe AD patients already have a A β level that is too high, resulting in plaques. The presence of plaques has shown to disable clearance⁹⁸, which could be the reason why supplementation of melatonin doesn't work in severe patients.

As for orexin, suppressing the orexin system should lead to an increase of sleep and thus possibly to an increase of A β clearance. Research has indicated that a dual orexin receptor antagonist decreased the A β level. Furthermore, knocking out the orexin gene also reduced A β accumulation. This effect could also be reversed by sleep deprivation¹⁵⁰. Again indicating that sleep deprivation can contribute to the stockpiling of A β .

Looking at the immune system, research shows an increase of microglia and astrocytes after chronic SD^{166,167}. This activation of microglia has also been seen in AD¹⁶⁶. In SD, the increased level of A β could possibly trigger the activation of microglial cells by self-aggregation^{99,169,170}.

Overall, it is fascinating to consider SD as a major risk factor so far. However, research does support the idea of SD as an influence on the development of AD, especially in people with a high risk at AD. To what extent SD influences the development is not well understood. What can be concluded is that normalization of sleep-wake patterns contribute to the clearance of metabolic waste in the brain and reduces the risk at AD. As for moderate AD patients, this potential therapy target could be of major influence on the prognosis of the disease. Considering, it is proven that patients with AD and SD are more likely to get symptomatic. To extent the impact of sleep treatment it is of importance to find a way to diagnose AD as early as possible.

6. References

1. Di Meco, A., Joshi, Y. B. & Praticò, D. Sleep deprivation impairs memory, tau metabolism, and synaptic integrity of a mouse model of Alzheimer's disease with plaques and tangles. *Neurobiol. Aging* **35**, 1813–1820 (2014).
2. Lucey, B. P. & Bateman, R. J. Amyloid- β diurnal pattern: Possible role of sleep in Alzheimer's disease pathogenesis. *Neurobiology of Aging* **35**, S29–S34 (2014).
3. Huang, Y. & Mucke, L. Alzheimer mechanisms and therapeutic strategies. *Cell* **148**, 1204–1222 (2012).
4. Miller, M. A. The role of sleep and sleep disorders in the development, diagnosis, and management of neurocognitive disorders. *Front. Neurol.* **6**, (2015).
5. Wimo, A., Jönsson, L., Bond, J., Prince, M. & Winblad, B. The worldwide economic impact of dementia 2010. *Alzheimer's Dement.* **9**, 1–11 (2013).
6. Prince, M. *et al.* The global prevalence of dementia: A systematic review and metaanalysis. *Alzheimer's and Dementia* **9**, 63–75 (2013).
7. Bates, K. a *et al.* Clearance mechanisms of Alzheimer's amyloid-beta peptide: implications for therapeutic design and diagnostic tests. *Mol. Psychiatry* **14**, 469–86 (2009).
8. Bodea, L. G., Eckert, A., Ittner, L. M., Piguet, O. & Götz, J. Tau physiology and pathomechanisms in frontotemporal lobar degeneration. *J. Neurochem.* **138**, 71–94 (2016).
9. Ballard, C. *et al.* Alzheimer's disease. *Lancet* **377**, 1019–1031 (2011).
10. Kepp, K. P. Alzheimer's disease due to loss of function: A new synthesis of the available data. *Prog. Neurobiol.* **143**, 36–60 (2016).
11. Lim, A. S. P. *et al.* Modification of the relationship of the apolipoprotein E $\epsilon 4$ allele to the risk of Alzheimer disease and neurofibrillary tangle density by sleep. *JAMA Neurol.* **70**, 1544–51 (2013).
12. Ju, Y.-E. S., Lucey, B. P. & Holtzman, D. M. Sleep and Alzheimer disease pathology—a bidirectional relationship. *Nat. Rev. Neurol.* **10**, 115–119 (2014).
13. Arendt, T., Stieler, J. & Holzer, M. Brain hypometabolism triggers PHF-like phosphorylation of tau, a major hallmark of Alzheimer's disease pathology. *Journal of Neural Transmission* **122**, 531–539 (2015).
14. Brettschneider, J., Del Tredici, K., Lee, V. & Trojanowski, J. Q. Spreading of pathology in neurodegenerative diseases: a focus on human studies. *Nat. Rev. Neurosci.* **16**, 190–120 (2015).
15. Dá, S. *et al.* Neuroscience and Biobehavioral Reviews Insights on the pathophysiology of Alzheimer's disease : The crosstalk between amyloid pathology , neuroinflammation and the peripheral immune system. *Neurosci. Biobehav. Rev.* **68**, 547–562 (2016).
16. Kumar, A. & Singh, A. A review on Alzheimer's disease pathophysiology and its management: an upda. *Pharmacol. Reports* **67**, 195–203 (2015).
17. Kurz, A. & Perneczky, R. Progress in Neuro-Psychopharmacology & Biological Psychiatry Novel insights for the treatment of Alzheimer's disease. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **35**, 373–379 (2011).

18. Chow, V. W., Mattson, M. P., Wong, P. C. & Gleichmann, M. Neuronal Activity and the Expression of Clathrin Assembly Protein AP180. *Biochem Biophys Res Commun* **12**, 1–12 (2011).
19. Querfurth, H. W. & LaFerla, F. M. Alzheimer's Disease. *N. Engl. J. Med.* **16**, 56–67 (2004).
20. Mawuenyega, K. G. *et al.* Decreased Clearance of CNS Amyloid-B in Alzheimer's Disease. *Science (80-.)*. **330**, 1774 (2010).
21. Leissring, M. A. *et al.* Enhanced proteolysis of beta-amyloid in APP transgenic mice prevents plaque formation, secondary pathology, and premature death. *Neuron* **40**, 1087–1093 (2003).
22. Mhyre, T. R., Boyd, J. T., Hamill, R. W. & Maguire-Zeiss, K. A. Protein Aggregation and Fibrillogenesis in Cerebral and Systemic Amyloid Disease. *Subcell. Biochem.* **65**, 389–455 (2012).
23. Fang, H., Zhang, L.-F., Meng, F.-T., Du, X. & Zhou, J.-N. *Acute hypoxia promote the phosphorylation of tau via ERK pathway.* *Neuroscience Letters* **474**, (2010).
24. Perry, E. K., Gibson, P. H., Blessed, G., Perry, R. H. & Tomlinson, B. E. Neurotransmitter enzyme abnormalities in senile dementia. Choline acetyltransferase and glutamic acid decarboxylase activities in necropsy brain tissue. *J. Neurol. Sci.* **34**, 247–265 (1977).
25. Nilsson, L., Nordberg, A., Hardy, J., Wester, P. & Winblad, B. Physostigmine restores 3H-acetylcholine efflux from Alzheimer brain slices to normal level. *J. Neural Transm.* **67**, 275–285 (1986).
26. Rylett, R. J., Ball, M. J. & Colhoun, E. H. Evidence for high affinity choline transport in synaptosomes prepared from hippocampus and neocortex of patients with Alzheimer's disease. *Brain Res.* **289**, 169–175 (1983).
27. Bowen, D. M., Smith, C. B., White, P. & Davison, A. N. Neurotransmitter-related enzymes and indices of hypoxia in senile dementia and other abiotrophies. *Brain* **99**, 459–496 (1976).
28. Davies, P. & Maloney, A. J. F. Selective loss of central cholinergic neurons in Alzheimer's disease. *Lancet* 1403 (1976).
29. Bartus, R. T., Dean, R. L., Beer, B. & Lippa, A. S. The cholinergic hypothesis of geriatric memory dysfunction. 408–417 (1982).
30. Drachman, D. A. Human Memory and the Cholinergic System. *Arch. Neurol.* **30**, 113 (1974).
31. Auld, D. S., Kornecook, T. J., Bastianetto, S. & Quirion, R. Alzheimer's disease and the basal forebrain cholinergic system: relations to beta-amyloid peptides, cognition, and treatment strategies. *Prog. Neurobiol.* **68**, 209–245 (2002).
32. Alvarez, A. *et al.* Stable complexes involving acetylcholinesterase and amyloid-beta peptide change the biochemical properties of the enzyme and increase the neurotoxicity of Alzheimer's fibrils. *J. Neurosci.* **18**, 3213–23 (1998).
33. Talesa, V. N. Acetylcholinesterase in Alzheimer's disease. in *Mechanisms of Ageing and Development* **122**, 1961–1969 (2001).
34. Becker, R. E., Moriarty, P. & Unni, L. in *The second generation of cholinesterase inhibitors: clinical and pharmacological effects.* 263–296 (1991).
35. Francis, P. T., Palmer, A. M., Snape, M. & Wilcock, G. K. The cholinergic hypothesis of Alzheimer's

- disease : a review of progress. *Journal Neurol. Neurosurg. Psychiatry* **66**, 137–147 (1999).
36. Cummings, J. L. *et al.* 'Metrifonate treatment of the cognitive deficits of Alzheimer's disease': Correction. *Neurology* (1998).
 37. Corey-Bloom, J., Anand, R. & Veach, J. A randomized trial evaluating the efficacy and safety of ENA 713 (rivastigmine tartrate), a new acetylcholinesterase inhibitor, in patients with mild to moderately severe Alzheimer's disease. *Int. J. Geriatr. Psychopharmacol.* (1998).
 38. Morris, J. C. *et al.* Metrifonate benefits cognitive, behavioral, and global function in patients with Alzheimer's disease [see comments]. *Neurology* (1998). doi:10.1212/WNL.50.5.1222
 39. Contestabile, A. The history of the cholinergic hypothesis. *Behav. Brain Res.* **221**, 334–340 (2011).
 40. Zhang, B. *et al.* Integrated systems approach identifies genetic nodes and networks in late-onset Alzheimer's disease. *Cell* **153**, 707–720 (2013).
 41. Heppner, F. L., Ransohoff, R. M. & Becher, B. Immune attack: the role of inflammation in Alzheimer disease. *Nat Rev Neurosci* **16**, 358–372 (2015).
 42. Stewart, W. F., Kawas, C., Corrada, M. & Metter, E. J. Risk of Alzheimer's disease and duration of NSAID use. *Neurology* **48**, 626–632 (1997).
 43. Sofroniew, M. V. & Vinters, H. V. Astrocytes: Biology and pathology. *Acta Neuropathologica* **119**, 7–35 (2010).
 44. Ullian, E. M. Control of Synapse Number by Glia. *Science (80-.)*. **291**, 657–661 (2001).
 45. Minagar, A. *et al.* The role of macrophage/microglia and astrocytes in the pathogenesis of three neurologic disorders: HIV-associated dementia, Alzheimer disease, and multiple sclerosis. *J. Neurol. Sci.* **202**, 13–23 (2002).
 46. Halassa, M. M. & Haydon, P. G. Integrated Brain Circuits: Astrocytic Networks Modulate Neuronal Activity and Behavior. *Annu. Rev. Physiol.* (2010). doi:10.1146/annurev-physiol-021909-135843
 47. Henneberger, C., Papouin, T., Oliet, S. H. R. & Rusakov, D. A. Long-term potentiation depends on release of d-serine from astrocytes. *Nature* (2010). doi:10.1038/nature08673
 48. Perea, G. *et al.* Tripartite synapses: astrocytes process and control synaptic information. *Trends Neurosci.* **32**, 421–31 (2009).
 49. Oberheim, N. A. *et al.* Loss of Astrocytic Domain Organization in the Epileptic Brain. *J. Neurosci.* **28**, 3264–3276 (2008).
 50. Neumann, H., Kotter, M. R. & Franklin, R. J. M. Debris clearance by microglia: An essential link between degeneration and regeneration. *Brain* **132**, 288–295 (2009).
 51. Nimmerjahn, A., Kirchhoff, F. & Helmchen, F. Resting microglial cells are highly dynamic surveillants of brain parenchyma in vivo. *Neuroforum* (2005). doi:10.1126/science.1110647
 52. Ransohoff, R. M. & Perry, V. H. Microglial Physiology: Unique Stimuli, Specialized Responses. *Annu. Rev. Immunol.* **27**, 119–145 (2009).
 53. Heneka, M. T. *et al.* Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* **14**, 388–405 (2015).

54. Lee, C. Y. D. & Landreth, G. E. The role of microglia in amyloid clearance from the AD brain. *Journal of Neural Transmission* **117**, 949–960 (2010).
55. Hickman, S. E., Allison, E. K. & El Khoury, J. Microglial Dysfunction and Defective β -Amyloid Clearance Pathways in Aging Alzheimer's Disease Mice. *J. Neurosci.* **28**, 8354–8360 (2008).
56. Prokop, S., Miller, K. R. & Heppner, F. L. Microglia actions in Alzheimer's disease. *Acta Neuropathologica* **126**, 461–477 (2013).
57. Hickman, S. E. *et al.* The microglial sensome revealed by direct RNA sequencing. *Nat. Neurosci.* **16**, 1896–1905 (2013).
58. El Khoury, J. B. *et al.* CD36 Mediates the Innate Host Response to β -Amyloid. *J. Exp. Med.* (2003). doi:10.1084/jem.20021546
59. Stewart, C. R. *et al.* CD36 ligands promote sterile inflammation through assembly of a Toll-like receptor 4 and 6 heterodimer. *Nat. Immunol.* (2010). doi:10.1038/ni.1836
60. Miller, M. A., Wright, H., Ji, C. & Cappuccio, F. P. Cross-sectional study of sleep quantity and quality and amnesic and non-amnesic cognitive function in an ageing population: the English Longitudinal Study of Ageing (ELSA). *PLoS One* **9**, e100991 (2014).
61. Craig, D., Hart, D. J. & Passmore, A. P. Genetically Increased Risk of Sleep Disruption in Alzheimer's Disease. *Sleep* **29**, 1003–1007 (2006).
62. Weber, F. & Dan, Y. Circuit-based interrogation of sleep control. *Nature* **538**, 51–59 (2016).
63. Boostani, R., Karimzadeh, F. & Torabi-Nami, M. A Comparative Review on Sleep Stage Classification Methods in Patients and healthy Individuals. *Comput. Methods Programs Biomed.* **140**, 77–91 (2016).
64. Gooneratne, N. S. & Vitiella, M. V. Sleep In Older Adults: Normative Changes, Sleep Disorders, and Treatment Options Nalaka. *Clin. Geriatr. Med.* **30**, 591–627 (2014).
65. Wolkove, N., Elkholy, O., Baltzan, M. & Palayew, M. Sleep and aging: 1. Sleep disorders commonly found in older people. *Review* **176**, 1299–1304 (2007).
66. Yoon, S. S. & Jo, S. A. Mechanisms of amyloid- β peptide clearance: Potential therapeutic targets for Alzheimer's disease. *Biomolecules and Therapeutics* **20**, 245–255 (2012).
67. Chesser, A. S., Pritchard, S. M. & Johnson, G. V. W. Tau clearance mechanisms and their possible role in the pathogenesis of Alzheimer disease. *Frontiers in Neurology* **4 SEP**, (2013).
68. Iliff, J. J. *et al.* Brain-wide pathway for waste clearance captured by contrast-enhanced MRI. *J. Clin. Invest.* **123**, 1299–1309 (2013).
69. Iliff, J. J. *et al.* A Paravascular Pathway Facilitates CSF Flow Through the Brain Parenchyma and the Clearance of Interstitial Solutes, Including Amyloid β . *Sci. Transl. Med.* **4**, (2012).
70. Iliff, J. J. & Nedergaard, M. Is there a cerebral lymphatic system? *Stroke* **601**, s93–s95 (2013).
71. Mendelsohn, A. R. & Larrick, J. W. Sleep Facilitates Clearance of Metabolites from the Brain: Glymphatic Function in Aging and Neurodegenerative Diseases. *Rejuvenation Res.* **16**, 518–523 (2013).

72. Jessen, N. A., Finmann Munk, A. S., Lundgaard, I. & Nedergaard, M. The Glymphatic System – A Beginner’s Guide Nadia. *Neurochem. Res.* **40**, 2583–2599 (2015).
73. Thrane, V. R. *et al.* Paravascular microcirculation facilitates rapid lipid transport and astrocyte signaling in the brain. *Sci. Rep.* **3**, (2013).
74. Murtha, L. a *et al.* Cerebrospinal fluid is drained primarily via the spinal canal and olfactory route in young and aged spontaneously hypertensive rats. *Fluids Barriers CNS* (2014). doi:10.1186/2045-8118-11-12
75. Johnston, M., Zakharov, A., Papaiconomou, C., Salmasi, G. & Armstrong, D. Evidence of connections between cerebrospinal fluid and nasal lymphatic vessels in humans, non-human primates and other mammalian species. *Cerebrospinal Fluid Res.* (2004). doi:10.1186/1743-8454-1-2
76. Xie, L. *et al.* Sleep Drives Metabolite Clearance from the Adult Brain. *Science (80-.)*. **342**, 373–377 (2013).
77. Stickgold, R. Neuroscience: A memory boost while you sleep. *Nature* **444**, 559–560 (2006).
78. Pace-Schott, E. F. & Spencer, R. M. C. *Age-related changes in the cognitive function of sleep. Progress in Brain Research* **191**, (Elsevier B.V., 2011).
79. Miller, M. A., Wright, H., Hough, J. & Cappuccio, F. P. in (ed. Idzikowski, C. B. T.-S. and its D. A. S.) Ch. 01 (InTech, 2014). doi:10.5772/58735
80. Salzarulo, P. *et al.* Sleep stages preceding spontaneous awakenings in the elderly. *Sleep Res. Online* (1999).
81. Dijk, D. J., Duffy, J. F. & Czeisler, C. a. Age-related increase in awakenings: impaired consolidation of nonREM sleep at all circadian phases. *Sleep* (2001).
82. Van Cauter, E., Leproult, R. & Plat, L. Age-related changes in slow wave sleep and REM sleep and relationship with growth hormone and cortisol levels in healthy men. *J. Am. Med. Assoc.* (2000).
83. Sowell ER *et al.* Mapping cortical change across the human life span. *Nat. Neurosci.* (2003). doi:10.1038/nn1008
84. Mander, B. A. *et al.* Prefrontal atrophy, disrupted NREM slow waves, and impaired hippocampal-dependent memory in aging. *Nat. Neurosci.* **16**, 357–364 (2013).
85. Mander, B. A. *et al.* β -amyloid disrupts human NREM slow waves and related hippocampus-dependent memory consolidation. *Nat. Neurosci.* **18**, 1051–1057 (2015).
86. Cricco, M., Simonsick, E. M. & Foley, D. J. The impact of insomnia on cognitive functioning in older adults. *J. Am. Geriatr. Soc.* **49**, 1185–1189 (2001).
87. Osorio, R. S. *et al.* Greater risk of Alzheimer’s disease in older adults with insomnia. *Journal of the American Geriatrics Society* **59**, 559–562 (2011).
88. Tranah, G. J. *et al.* Circadian activity rhythms and risk of incident dementia and mild cognitive impairment in older women. *Ann. Neurol.* **70**, 722–732 (2011).
89. Hahn, E. A., Wang, H. X., Andel, R. & Fratiglioni, L. A change in sleep pattern may predict alzheimer disease. *Am. J. Geriatr. Psychiatry* **22**, 1262–1271 (2014).

90. Lim, A. S. P., Kowgier, M., Yu, L., Buchman, A. S. & Bennett, D. A. Sleep Fragmentation and the Risk of Incident Alzheimer's Disease and Cognitive Decline in Older Persons. *Sleep* **36**, 1027–1032 (2013).
91. Yaffe, K. *et al.* Sleep-Disordered Breathing, Hypoxia, and Risk of Mild Cognitive Impairment and Dementia in Older Women. *JAMA J. Am. Med. Assoc.* **306**, 613 (2011).
92. Spira, A. P. *et al.* Self-reported sleep and beta-amyloid deposition in community-dwelling older adults. *JAMA Neurol* **70**, 1537–1543 (2013).
93. Harper, D. G. *et al.* Dorsomedial SCN neuronal subpopulations subserve different functions in human dementia. *Brain* **131**, 1609–1617 (2008).
94. Hita-Yanez, E., Atienza, M. & Cantero, E. G.-N. and J. L. Disturbed Sleep Patterns in Elders with Mild Cognitive Impairment: The Role of Memory Decline and ApoE ε4 Genotype. *Current Alzheimer Research* **9**, 290–297 (2012).
95. Moran, M. *et al.* Sleep disturbance in mild to moderate Alzheimer's disease. *Sleep Med.* **6**, 347–352 (2005).
96. Wisor, J. P. *et al.* Sleep and circadian abnormalities in a transgenic mouse model of Alzheimer's disease: A role for cholinergic transmission. *Neuroscience* **131**, 375–385 (2005).
97. Cirrito, J. R. *et al.* Synaptic activity regulates interstitial fluid amyloid-beta levels in vivo. *Neuron* **48**, 913–922 (2005).
98. Huang, Y., Potter, R., Sigurdson, W. & al, et. Effects of age and amyloid deposition on aβ dynamics in the human central nervous system. *Arch. Neurol.* **69**, 51–58 (2012).
99. Kang, J.-E. *et al.* Amyloid-B Dynamics Are Regulated by Orexin and the Sleep-Wake Cycle. *Science* (80-.). **326**, 1005–1007 (2009).
100. Ooms, S. *et al.* Effect of 1 Night of Total Sleep Deprivation on Cerebrospinal Fluid β-Amyloid 42 in Healthy Middle-Aged Men: A Randomized Clinical Trial. *JAMA Neurol.* **71**, 971–977 (2014).
101. Palma, J.-A., Urrestarazu, E. & Iriarte, J. Sleep loss as risk factor for neurologic disorders: a review. *Sleep Med.* **14**, 229–36 (2013).
102. YS, J., JS, M., CD, T. & al, et. Sleep quality and preclinical alzheimer disease. *JAMA Neurol.* **70**, 587–593 (2013).
103. Cantero, J. L. *et al.* Tau protein role in sleep-wake cycle. *J. Alzheimer's Dis.* **21**, 411–421 (2010).
104. Jarzynka, M. J. *et al.* Microtubules modulate melatonin receptors involved in phase-shifting circadian activity rhythms: In vitro and in vivo evidence. *J. Pineal Res.* **46**, 161–171 (2009).
105. Jarzynka, M. J. *et al.* Modulation of melatonin receptors and G-protein function by microtubules. *J. Pineal Res.* **41**, 324–336 (2006).
106. Holth, J. K., Mahan, T. E., Robinson, G. O., Rocha, A. & Holtzman, D. M. Altered sleep and EEG power in the P301S Tau transgenic mouse model. *Ann. Clin. Transl. Neurol.* 180–190 (2017). doi:10.1002/acn3.390
107. Wang, Y. & Mandelkow, E. Tau in physiology and pathology. *Nat Rev Neurosci* **17**, 22–35 (2016).

108. Yamada, K. *et al.* Neuronal activity regulates extracellular tau in vivo. *J. Exp. Med.* **211**, 387–393 (2014).
109. Liao, X., Zhang, Y., Wang, Y. & Wang, J. The effect of cdk-5 overexpression on tau phosphorylation and spatial memory of rat. *Sci. China Ser. C Life Sci.* **47**, 251–257 (2004).
110. Tsai, L.-H., Lee, M.-S. & Cruz, J. Cdk5, a therapeutic target for Alzheimer's disease? *Biochim. Biophys. Acta - Proteins Proteomics* **1697**, 137–142 (2004).
111. Rothman, S. M., Herdener, N., Frankola, K. A., Mughal, M. R. & Mattson, M. P. Chronic mild sleep restriction accentuates contextual memory impairments, and accumulations of cortical A β and pTau in a mouse model of Alzheimer's disease. *Brain Res.* (2013). doi:10.1016/j.brainres.2013.07.010
112. Spiegel, K., Knutson, K., Leproult, R., Tasali, E. & Van Cauter, E. Sleep loss: A novel risk factor for insulin resistance and Type 2 diabetes. *J. Appl. Physiol.* **99**, 2008–2019 (2005).
113. Donga, E. *et al.* A single night of partial sleep deprivation induces insulin resistance in multiple metabolic pathways in healthy subjects. *J. Clin. Endocrinol. Metab.* **95**, 2963–2968 (2010).
114. Ott, A. *et al.* Diabetes mellitus and the risk of dementia: The Rotterdam Study. *Neurology* **53**, 1937–42 (1999).
115. Arvanitakis, Z., Wilson, R. S., Bienias, J. L., Evans, D. A. & Bennett, D. A. Diabetes mellitus and risk of Alzheimer disease and decline in cognitive function. *Arch. Neurol.* **61**, 661–666 (2004).
116. Murakami, K. *et al.* Insulin receptor mutation results in insulin resistance and hyperinsulinemia but does not exacerbate Alzheimer's-like phenotypes in mice. *Biochem. Biophys. Res. Commun.* **409**, 34–39 (2011).
117. Henderson, A. S. *et al.* Apolipoprotein E allele epsilon 4, dementia, and cognitive decline in a population sample. *Lancet* (1995). doi:10.1016/S0140-6736(95)92405-1
118. Genin, E. *et al.* ApoE and Alzheimer disease: a major gene with semi-dominant inheritance. *Mol. Psychiatry* **16**, 903–907 (2011).
119. Hita-Yañez, E., Atienza, M. & Cantero, J. L. Polysomnographic and Subjective Sleep Markers of Mild Cognitive Impairment. *Sleep* **36**, 1327–1334 (2013).
120. Kadotani, H., Kadotani, T., Young, T. & al, et. Association between apolipoprotein e ϵ 4 and sleep-disordered breathing in adults. *JAMA* **285**, 2888–2890 (2001).
121. Liu, R.-Y., Zhou, J.-N., van Heerikhuizen, J., Hofman, M. A. & Swaab, D. F. Decreased Melatonin Levels in Postmortem Cerebrospinal Fluid in Relation to Aging, Alzheimer's Disease, and Apolipoprotein E- ϵ 4/4 Genotype1. *J. Clin. Endocrinol. Metab.* **84**, 323–327 (1999).
122. Gottlieb, D. J. *et al.* APOE ϵ 4 is associated with obstructive sleep apnea/hypopnea: The Sleep Heart Health Study . *Neurol.* **63**, 664–668 (2004).
123. Kaushal, N., Ramesh, V. & Gozal, D. Human apolipoprotein E4 targeted replacement in mice reveals increased susceptibility to sleep disruption and intermittent hypoxia. *Am. J. Physiol. - Regul. Integr. Comp. Physiol.* **303**, R19 LP-R29 (2012).
124. O'Hara, R. *et al.* Nocturnal sleep apnea/hypopnea is associated with lower memory performance

- in APOE ϵ 4 carriers. *Neurol.* **65**, 642–644 (2005).
125. Spira, A. P. *et al.* Sleep-disordered breathing and cognition in older women. *J. Am. Geriatr. Soc.* **56**, 45–50 (2008).
 126. Yesavage, J. A. *et al.* Sleep/Wake Disruption in Alzheimer's Disease: APOE Status and Longitudinal Course. *J. Geriatr. Psychiatry Neurol.* **17**, 20–24 (2004).
 127. Spira, A. P. *et al.* Objectively measured sleep and β -amyloid burden in older adults: A pilot study. *SAGE Open Med.* **2**, 2050312114546520 (2014).
 128. Cosentino, F. I. I. *et al.* The APOE epsilon4 allele increases the risk of impaired spatial working memory in obstructive sleep apnea. *Sleep Med.* **9**, 831–839 (2008).
 129. Lin, L. *et al.* Melatonin in Alzheimer's disease. *Int. J. Mol. Sci.* **14**, 14575–14593 (2013).
 130. Pappolla, M. a *et al.* An assessment of the antioxidant and the antiamyloidogenic properties of melatonin: implications for Alzheimer's disease. *J. Neural Transm.* **107**, 203–231 (2000).
 131. Rosales-Corral, S. A. *et al.* Alzheimer's disease: pathological mechanisms and the beneficial role of melatonin. *J. Pineal Res.* **52**, 167–202 (2012).
 132. Tohgi, H. *et al.* Concentrations of serotonin and its related substances in the cerebrospinal fluid in patients with Alzheimer type dementia. *Neurosci. Lett.* **141**, 9–12 (1992).
 133. Mishima, K. *et al.* Melatonin secretion rhythm disorders in patients with senile dementia of Alzheimer's type with disturbed sleep–waking. *Biol. Psychiatry* **45**, 417–421 (1999).
 134. Magri, F. *et al.* Changes in endocrine circadian rhythms as markers of physiological and pathological brain aging. *Chronobiol. Int.* **14**, 385–396 (1997).
 135. Gehrman, P. R. *et al.* Melatonin Fails To Improve Sleep Or Agitation In A Double-Blind Randomized Placebo-Controlled Trial Of Institutionalized Patients With Alzheimer's Disease. *Am. J. Geriatr. Psychiatry* **17**, 166–169 (2009).
 136. Singer, C. *et al.* A Multicenter, Placebo-controlled Trial of Melatonin for Sleep Disturbance in Alzheimer's Disease. *Sleep* **26**, 893–901 (2003).
 137. Brzezinski, A. *et al.* Effects of exogenous melatonin on sleep: a meta-analysis. *Sleep Med. Rev.* **9**, 41–50 (2005).
 138. Cardinali, D. P., Brusco, L., Liberczuk, C. & Furio, A. M. The use of melatonin in Alzheimer's disease. *Neuro Endocrinol. Lett.* **23**, 20–23 (2002).
 139. Brusco, L., Márquez, M. & Cardinali, D. P. Melatonin treatment stabilizes chronobiologic and cognitive symptoms in Alzheimer's disease. *Neuro Endocrinol. Lett.* **21**, 39–42 (2000).
 140. Mishima, K., Okawa, M., Hozumi, S. & Hishikawa, Y. Supplementary administration of artificial bright light and melatonin as potent treatment for disorganized circadian rest-activity and dysfunctional autonomic and neuroendocrine systems in institutionalized demented elderly persons. *Chronobiol. Int.* **17**, 419–432 (2000).
 141. Brusco, L. I., Márquez, M. & Cardinali, D. P. Monozygotic twins with Alzheimer's disease treated with melatonin: Case report. *J. Pineal Res.* **25**, 260–263 (1998).

142. Lahiri, D. K. Melatonin affects the metabolism of the β -amyloid precursor protein in different cell types. *J. Pineal Res.* **26**, 137–146 (1999).
143. Matsubara, E. *et al.* Melatonin increases survival and inhibits oxidative and amyloid pathology in a transgenic model of Alzheimer's disease. *J. Neurochem.* **85**, 1101–1108 (2003).
144. Zhang, Y. C., Wang, Z. F., Wang, Q., Wang, Y. P. & Wang, J. Z. Melatonin attenuates beta-amyloid-induced inhibition of neurofilament expression. *Acta Pharmacol. Sin.* **25**, 447–451 (2004).
145. P. Cardinali, D., M. Furio, A. & I. Brusco, L. Clinical Aspects of Melatonin Intervention in Alzheimers Disease Progression. *Curr. Neuropharmacol.* **8**, 218–227 (2010).
146. Monti, J. M., Alvariño, F., Cardinali, D., Savio, I. & Pintos, A. Polysomnographic study of the effect of melatonin on sleep in elderly patients with chronic primary insomnia. *Arch. Gerontol. Geriatr.* **28**, 85–98 (2017).
147. Wade, A. G. *et al.* Add-on prolonged-release melatonin for cognitive function and sleep in mild to moderate Alzheimer's disease: a 6-month, randomized, placebo-controlled, multicenter trial. *Clin. Interv. Aging* **9**, 947–961 (2014).
148. Peter-Derex, L., Yammine, P., Bastuji, H. & Croisile, B. Sleep and Alzheimer's disease. *Sleep Medicine Reviews* **19**, 29–38 (2015).
149. Fronczek, R. *et al.* Hypocretin (orexin) loss in Alzheimer's disease. *Neurobiol. Aging* **33**, 1642–1650 (2011).
150. Roh, J. H. *et al.* Potential role of orexin and sleep modulation in the pathogenesis of Alzheimer's disease. *J. Exp. Med.* **211**, 2487–96 (2014).
151. Keene, A. C. & Joiner, W. J. Neurodegeneration: Paying it off with sleep. *Curr. Biol.* **25**, R234–R236 (2015).
152. Scammell, T. E., Matheson, J. K., Honda, M., Thannickal, T. C. & Siegel, J. M. Coexistence of narcolepsy and Alzheimer's disease. *Neurobiol. Aging* **33**, 1318–1319 (2012).
153. Savard, J., Laroche, L., Simard, S., Ivers, H. & Morin, M. M. Chronic Insomnia and Immune Functioning. *Psychosom. Med.* **221**, 211–221 (2003).
154. Irwin, M. R., Witarana, T., Caudill, M., Olmstead, R. & Breen, E. C. Brain , Behavior , and Immunity Sleep loss activates cellular inflammation and signal transducer and activator of transcription (STAT) family proteins in humans. *Brain, Behav. Immun.* (2014). doi:10.1016/j.bbi.2014.09.017
155. Shearer, W. T. *et al.* Soluble TNF- α receptor 1 and IL-6 plasma levels in humans subjected to the sleep deprivation model of spaceflight. *J. allergy Clin. Immunol.* 165–170 (2001). doi:10.1067/mai.2001.112270
156. Banks, W. A., Kastin, A. J. & Broadwell, R. D. Passage of Cytokines across the blood-brain barrier. *Neuroimmunomodulation* **2**, 241–248 (1995).
157. Bryant, P. A., Trinder, J. & Curtis, N. SICK AND TIRED: DOES SLEEP HAVE A VITAL ROLE IN THE IMMUNE SYSTEM ? *Nat. Rev. Immunol.* **4**, 457–467 (2004).
158. Besedovsky, L., Lange, T. & Born, J. Sleep and immune function. *Pflugers Arch. Eur. J. Physiol.* **463**, 121–137 (2012).

159. Moldofsky, H., Lue, F. A., Davidson, J. R. & Gorczynski, R. Effects of sleep deprivation on human immune functions. *FASEB J.* **3**, 1972–7 (1989).
160. Brambilla, D., Franciosi, S., Opp, M. R. & Imeri, L. Interleukin-1 inhibits firing of serotonergic neurons in the dorsal raphe nucleus and enhances GABAergic inhibitory post-synaptic potentials. *Eur. J. Neurosci.* **26**, 1862–1869 (2007).
161. Krueger, J. M., Obál, F., Fang, J., Kubota, T. & Taishi, P. The Role of Cytokines in Physiological Sleep Regulation. *Ann. N. Y. Acad. Sci.* **933**, 211–221 (2001).
162. Opp, M. R. Cytokines and sleep. *Sleep Med. Rev.* **9**, 355–364 (2005).
163. Olivadoti, M. D. & Opp, M. R. Effects of intracerebroventricular administration of interleukin (IL)-1 on sleep and body temperature of IL-6-deficient mice. *Neuroscience* **153**, 338–348 (2008).
164. Lancel, M., Mathias, S., Faulhaber, J., Schiffelholz, T. & Mathias, S. Effect of interleukin-1 beta on EEG power density during sleep depends on circadian phase. *Am J Physiol Regul Integr Comp Physiol* **270**, R830–R837 (1996).
165. Opp, M. R., Obal, F. & Krueger, J. M. Interleukin 1 alters rat sleep: temporal and dose-related effects. *Am. J. Physiol. - Regul. Integr. Comp. Physiol.* **260**, R52 LP-R58 (1991).
166. Bellesi, X. M. *et al.* Sleep Loss Promotes Astrocytic Phagocytosis and Microglial Activation in Mouse Cerebral Cortex. *J. Neurosci.* **37**, 5263–5273 (2017).
167. Hsu, J., Lee, Y., Chang, C., Ling, E. & Lan, C. Sleep deprivation prior to transient global cerebral ischemia attenuates glial reaction in the rat hippocampal formation. *Brain Res.* **984**, 170–181 (2003).
168. Bellesi, M., de Vivo, L., Tononi, G. & Cirelli, C. Effects of sleep and wake on astrocytes: clues from molecular and ultrastructural studies. *BMC Biol.* **13**, 66 (2015).
169. Halle, A. *et al.* The NALP3 inflammasome is involved in the innate immune response to amyloid- β . *Nat Immunol* **9**, 857–865 (2008).
170. Jung, C. K. E., Keppler, K., Steinbach, S., Blazquez-Llorca, L. & Herms, J. Fibrillar amyloid plaque formation precedes microglial activation. *PLoS One* **10**, 1–10 (2015).