



# PLASMA BIOMARKERS: A POSSIBLE WAY TO PREDICT EFFECTIVITY OF PD-1 BLOCKADE THERAPY IN HODGKIN LYMPHOMA

Jet Neervoort, S3481867 03-04-2020

Supervisor: prof. dr. J.H.M van den Berg Co-supervisor: dr. Lydia Visser Supervising PhD student: Johanna Veldman

Word count: 5757

# Content

Abstract	2
Introduction	2
Hodgkin lymphoma	4
Cause of Hodgkin lymphoma	4
Hodgkin lymphoma biology	4
Microenvironment	5
Current treatment	6
Immunotherapy	7
PD-1-PD-L1 expression in solid tumors	8
sPD-L1	8
sPD-L2	8
sPD-1	8
IDO	9
CD83	9
VEGF	9
PD-1-PD-L1 expression in Hodgkin lymphoma	. 10
sPD-L1	. 10
sPD-1	. 10
IDO	. 10
CD83	. 11
VEGF	. 11
Discussion	. 12
Conflict of interest	. 13
Acknowledgements	. 14
References	. 15

# Abstract

Hodgkin lymphoma is one of the most frequent lymphomas in the Western world and is characterized by an extremely low number of tumor cells within a large microenvironment. Treatment of Hodgkin lymphoma by radiotherapy and chemotherapy is very effective, but is highly toxic, causing second malignancies to be the leading cause of death among Hodgkin lymphoma survivors. Partly because of the large microenvironment surrounding the tumor cells, immune checkpoint blockade therapy targeting the PD-1/PD-L1 axis has shown great efficacy in relapsed and refractory Hodgkin lymphoma patients. Selection for potential responders to immunotherapy is very important and can ben realized by determination of predictive soluble biomarkers. The aim of this thesis is to identify the potential prognostic and predictive value of sPD-L1, sPD-L2, sPD-1, Indoleamine 2,3-dioxygenase (IDO), sCD83, and VEGF for PD-1 blockade therapy, and the possible impact of these soluble biomarkers on the PD-1/PL-L1 axis in Hodgkin lymphoma. We found that sPD-L1 and sPD-1 are promising independent predictive biomarkers and combination PD-1 blockade therapy with anti-IDO or anti-VEGF antibodies might provide a solution for resistance to immune checkpoint inhibition.

### Introduction

Over the past years, immune checkpoint therapy has become an important part of cancer treatment in both solid tumors and hematologic malignancies. Immune checkpoint therapy makes use of the intrinsic antitumor activity of the immune system through induction of an immune response to the tumor. Immune 'checkpoints', including cytotoxic T-lymphocyte-associated antigen-4 (CTLA-4) and the inhibitory receptor programmed cell death-1 (PD-1), normally function as inhibitory feedback loops to diminish immune activation. These checkpoints protect the host from autoimmunity or collateral tissue damage as a result of inflammation. This thesis mainly focuses on PD-1.

Programmed death-ligand 1 (PD-L1) is a membrane-bound protein primarily expressed on dendritic cells (DC) and monocytes (1,2). This ligand binds to the PD-1, expressed on activated T cells and B cells, DC, and monocytes (2). Research has shown that membrane-bound PD-1 and PD-L1 also have soluble forms, which will be discussed later (3). Upregulation of PD-L1 on antigen presenting cells can be accomplished when T cells recognize an antigen presented by human leukocyte antigen (HLA) molecules, which leads to the production of inflammatory cytokines (e.g. IFN-γ).

Cancer cells are not automatically recognized as 'foreign' by the immune system. Several mechanisms, such as regulatory immune cells, immunosuppressive cytokines and chemokines, and so-called 'immune-check-point' pathways down-modulate immune functions, resulting in a tumor-tolerant environment (4). This is frequently observed in a variety of cancers (5–8). Tumors subvert the immune response by increasing inhibitory immune checkpoint ligands like PD-L1 in the tumor microenvironment (TME), and therefore preventing an antitumor immune response through host T cell exhaustion (9). The PD-1-PD-L1 receptor-ligand pair serves as a dominant immune checkpoint pathway operative in the TME and regulates the strength of the immune response (10). PD-L1 expression is associated with worse clinical outcomes in several cancer types (1,11,12).

Increasing knowledge about the self-protecting mechanisms of tumor cells has led to the development of anti-PD-1 drugs like nivolumab and pembrolizumab. Monoclonal antibodies (mAbs) block the immune checkpoint pathways and therefore induce antitumor immune responses. Blockade of the PD-1 signaling pathway prevents inhibition of the immune response, allowing for the exhausted T cells to be reinvigorated

and induce an effective immune response, despite elevated levels of PD-L1 in the TME (10). This has appeared to be very effective in Hodgkin lymphoma, advanced melanoma, renal cell carcinoma, and non-small-cell lung cancer (NSCLC) (4,13,14). However, clinical success of this treatment greatly varies between different cancer types and different patients. Objective responses of 65-87% were achieved in Hodgkin lymphoma, whereas responses of 30% were reached in other malignancies (13).

Differences in response rates to immune checkpoint blockade might be explained by resistance. This phenomenon can be divided into two groups: 1) tumors that do not respond to therapy (primary resistance) and 2) tumors that initially respond, but eventually become resistant (acquired resistance) (13). Current knowledge about the mechanisms of resistance is limited, but Veldman, Visser, van den Berg, & Diepstra (13) described six potential mechanisms of PD-1 blockade resistance in Hodgkin lymphoma. 1) Shaping of a TME with downregulation of CD8+ T cells and upregulation of Treg cells (primary resistance); 2) Insufficient attraction and activation of T cells as a result of poor antigen presentation. Tumors with a high tumor mutation burden (TMB) present more neoantigens and as a result, are more sensitive to immune checkpoint blockade (15–17). Solid tumors are often characterized by a great number of genetic and epigenetic alterations, which leads to the presentation of a diverse set of neoantigens (18,19). (primary resistance); 3) elevated IDO levels, resulting in inhibited activation and function of effector T cells and leading to attraction and activation of preexisting Tregs and differentiation of naïve T cells into Tregs (primary resistance); 4) attraction of tumor associated macrophages (TAMs), which, in combination with PD-L1, can inhibit PD-1 + T cells and natural killer (NK) cells (primary resistance); 5) upregulation of PD-1, LAG-3 and TIM-3 by PD-1 blockade therapy (acquired resistance) and 6) deregulated purinergic signaling as a result of increased levels of adenosine. Adenosine is an immunosuppressive molecule that suppresses effector T cells and upregulates Treg in the TME (13).

As mentioned before, like numerous costimulatory molecules in immune regulation pathways, PD-1 and PD-L1 have membrane-bound forms and soluble forms. This increases the complexity and diversity of the PD-1-PD-L1 signaling pathway (3). However, they are a topic of interest in view of PD-L1 as a predictive biomarker because detection of biomarkers in plasma is easier and less invasive for the patient compared to a biopsy. The soluble form of PD-1 and PD-L1 is usually generated by proteolytic cleavage of the membrane-bound form of the molecule, or by translation of alternative spliced mRNA. There is growing evidence that sPD-1/PD-L1 might be as valuable as a predictive biomarker for clinical outcome as membrane-bound PD-1/PD-L1 (3). Tumor-related membrane-bound PD-1/PD-L1 contribute to immunosuppression. However, binding of sPD-1 to membrane-bound PD-1/PD-L2 may counteract the inhibitory effects of membrane-bound PD-1 on T cells by preventing the T cell inhibition as a result of membrane-bound PD-1-PD-L1/PD-L2 interaction (3,20). Whether higher levels of sPD-1 are associated with a better prognosis remains rather unclear (3,21–25).

Biomarkers including IDO, CD83, Vascular Endothelial Growth Factor (VEGF), IFN- $\gamma$ , and oncogene pathways such as IFN-  $\gamma$ /JAK2/IFN and PI3K might be involved in the generation of sPD-1 or sPD-L1 in patients, but the exact mechanisms by which this happens remain poorly understood (1,3). sPD-L1 has shown to be a meaningful and practical dynamic biomarker in several solid tumors such as melanoma (1), bladder cancer and NSCLC (4,26,27). IDO is an important immunosuppressive enzyme and expression can be induced by IFN- $\gamma$  (13). IDO is an important enzyme and the rate-limiting step in the kynurenine pathway. Through this pathway, T cell apoptosis is induced by degradation of tryptophan and increase of products from the kynurenine pathway (13,28), which leads to the conversion of naïve CD4+ T cells into Tregs (29). IDO has also been described as a predictive biomarker of anti-tumor response by immune checkpoint inhibitors. Expression of recombinant soluble CD83 (sCD83) downregulates DC maturation as well as DC dependent T cell responses. Thus, CD83 has an immunosuppressive function (30). The production of sCD83 is mostly mediated by activated B-cells and DC (31). VEGF is a proangiogenic signal protein but is also closely involved with induction of a immunosuppressive TME in cancer. In the TME, VEGF inhibits DC maturation, and causes induction of Treg cells. VEGF increases the expression levels of several immune checkpoints including PD-1, CTLA-4, TIM-3 and LAG3 on CD8+ T cells in the microenvironment, which results in T cell exhaustion (13,32,33).

Biomarkers can be used to select for patients with a higher likelihood to respond to immunotherapy. However, the role of several biomarkers in resistance to immune checkpoint therapy and the PD-1/PD-L1 axis in Hodgkin lymphoma is not yet clear. The aim of this study is to assess the influence of sPD-L1, sPD-L2, sPD-1, IDO, sCD83 and VEGF as biomarkers for the prediction of durable efficacy to immunotherapeutic agents and the PD-1-PD-L1 axis in Hodgkin lymphoma. To this end, we will summarize the current knowledge about the role of the TME in Hodgkin lymphoma, the mechanisms by which HRS cells evade the immune system and the impact of soluble markers including sPD-L1, sPD-L2, sPD-1, IDO, sCD83, and VEGF, on PD-1/PD-L1 expression in solid tumors as well as Hodgkin lymphoma.

### Hodgkin lymphoma

#### Cause of Hodgkin lymphoma

According to the American Cancer Society, approximately 8,480 new patients will be affected by Hodgkin lymphoma in the United States in 2020, making this disease one of the most frequent lymphomas in the Western world (34,35). The disease has an increased incidence in young adults (age 20-34) and patients 55 years and older. Risk factors for the development as well as the cause of the disease remain poorly understood. However, research suggests that an abnormal immune response to infection as a result of expression of a variety of cytokines and chemokines might play a role in the development of Hodgkin lymphoma (36). Epidemiologic and serologic studies suggest Epstein-Barr virus (EBV) in the cause of Hodgkin lymphoma (37). EBV positivity also influences the expression of certain cytokines including CXCL10, CXCL9 and CCL5, resulting in amplification of the intense inflammatory reaction to infection (36,38). However, other childhood infectious diseases including chickenpox, measles and rubella are associated with a decreased risk of Hodgkin lymphoma (39,40).

#### Hodgkin lymphoma biology

Hodgkin lymphoma is a B cell-lymphoid malignancy that can be subdivided into two distinct entities; classical Hodgkin lymphoma (around 95% of cases) and nodular lymphocyte predominant Hodgkin lymphoma (NLPHL; 5% of cases), based on morphological and clinical differences (13). Classical Hodgkin lymphoma is characterized by the presence of Hodgkin Reed-Sternberg (HRS) cells, while NLPHL is accompanied by the presence of lymphocyte-predominant (LP) cells (36,41). HRS cells are derived from germinal center B cells, but differ because they lack most B cell markers.

CD30 and CD15 are cell membrane proteins of the tumor necrosis factor (TNF)-receptor family. Under normal conditions, CD30 is not expressed in human tissue. However, co-expression of these membrane

proteins is typically associated with classical Hodgkin lymphoma (13,42–44). The presence of CD15 and CD30 on HRS cells promote tumor growth and survival. Therefore, these factors are interesting targets for cancer treatment (45).

Hodgkin lymphoma is a unique type of cancer because the amount of malignant cells is very low compared to the amount of reactive cells in a TME that includes lymphocytes, macrophages, eosinophils, mast cells, plasma cells, stromal cells, fibroblasts and other cells (36,46). In fact, the Hodgkin lymphoma tumor cell mass consists of only about 1% HRS cells (13). Therefore, the microenvironment is an important factor in the progression of the disease.

#### Microenvironment

Infiltrating CD4+ T cells, directly surrounding the HRS cells, are the most abundant cells found in the microenvironment of the tumor (36,47). Research shows that these cells primarily consist of T helper 2 (TH2) and T regulatory (Treg) cells (48,49). HRS cells express characteristic surface molecules and secrete a variety of cytokines and chemokines, resulting in interactions that shape the reactive infiltrate and signaling for the HRS. An overview of the interacting cells in the TME of Hodgkin lymphoma is shown in Fig 1. In Hodgkin lymphoma, several chemokines including CCL17 and CCL22 promote the attraction of CCR4expressing TH2 and Treg cells (50-54). CCL5 is a chemokine, commonly found in Hodgkin lymphoma, that has been shown to attract mast cells to the TME (55). Several other mechanisms, like the production of interferon gamma by HRS cells together with T helper 1 (TH1) cells (56,57), and the production of granulocyte colony stimulating factor (CSF1) and fractalkine (CX3CL1), chemokines and differentiation factors of monocytes by HRS cells, enhance the attraction and infiltration of macrophages (58-60). This is important for the progression of several cancers, including Hodgkin lymphoma, since macrophages promote cell migration and suppress antitumor immunity. Moreover, the secretion of macrophage migration inhibitory factor (MIF) by M2 macrophages may contribute to the proliferation of HRS cells (35,36). Activation and proliferation of fibroblasts in Hodgkin lymphoma is mediated by the expression of IL-13, tumor necrosis factor (TNF) α, and fibroblast growth factors by HRS cells (61). This, in turn, contributes to eosinophil and TH2 cell infiltration by secretion of CCL11 (35,62). Altogether, many cytokines and chemokines are present in the TME of Hodgkin lymphoma, which results in the attraction of cells that help the tumor survive. The complex interference of HRS cells with its microenvironment may provide novel strategies for cancer therapy.



**Fig 1. The tumor microenvironment in classical Hodgkin lymphoma.** A malignant Hodgkin Reed-Sternberg (HRS) cell expressing characteristic surface molecules as well as secreted cytokines and chemokines. These molecules attract the surrounding non-neoplastic cells. The arrows indicate the main activating and inhibitory functions of predominantly membrane-bound (purple triangles) and secreted molecules (pink), which are mediated by surface receptors. Not all existing interactions are shown. IL, interleukin; FGF, fibroblast growth factor; TNF- $\alpha$ , tumor necrosis factor  $\alpha$ ; LT- $\alpha$ , lymphotoxin  $\alpha$ ; TGF- $\beta$ , transforming growth factor  $\beta$ ; BAFF, B-cell activating factor; APRIL, a proliferation-inducing ligand; BLC, B lymphocyte chemoattractant; TNFR, tumor necrosis factor receptor; FASL, Fas ligand; MEC, mucosae-associated epithelial chemokine; Treg, T regulatory cell; TH, T helper cell; TARC, thymus and activation-related chemokine; MDC, Macrophage-derived chemokine; Gal-1, galectin-1; PDL, programmed cell death 1 ligand; IFN- $\gamma$ , interferon gamma; NK, natural killer (Steidl, Connors, & Gascoyne, 2011; (36)).

#### Current treatment

Over the last 60 years, there have been enormous improvements in the outlook for patients with Hodgkin lymphoma. Today, it is considered one of the most curable types of cancer because of the use of chemotherapy, sometimes in combination with radiotherapy. However, since the patient group is relatively young, this success has exposed the problem of long-term treatment-related toxicity. In fact, second malignancies are the leading cause of death among Hodgkin lymphoma survivors (63,64). Chemotherapy and radiotherapy are highly toxic, increasing the risk of several cancers, including breast cancer, lung cancer and acute myeloid leukemia (64). Furthermore, there is increased morbidity and mortality from ischaemic heart disease, valvular heart disease, anthracycline-related cardiomyopathy and conduction abnormalities among patients previously treated with radiotherapy (65,66). Altogether, new, less toxic therapy is highly

needed for Hodgkin lymphoma patients. Two essentials of new therapy are the minimization of long-term treatment-related toxicities and the improvement in therapy for patients with relapsed and refractory disease (19).

#### Immunotherapy

Immune checkpoints can be utilized to modulate the duration and amplitude of the physiological immune response (6,19). In solid tumors, including advanced melanoma and NSCLC, immune checkpoint inhibition has already shown great clinical success (4,13). Hodgkin lymphoma has a low mutational burden, resulting in a lower level of neoantigens. This could be an implication for immunotherapy in Hodgkin lymphoma patients (19,67). However, the amount of T cells, cytokines and chemokines present in the Hodgkin lymphoma microenvironment does make immune checkpoints an interesting target of therapy and Hodgkin lymphoma stands out among all lymphomas because of its high responsiveness to PD-1 blockade (>70% ORR) therapy and significant clinical benefit in patients with relapsed and refractory Hodgkin lymphoma (19,68–70).

Research has shown that Hodgkin patients often show deficiencies of host immune function, and patients with HIV infection or autoimmune disease have an increased risk of both Hodgkin lymphoma and non-Hodgkin lymphoma (19,71). Additionally, both PD-1 and its ligand PD-L1 are overexpressed in the TME of Hodgkin lymphoma patients, which contributes to a potent inhibitory signal in the maintenance of the immunosuppressive TME (19,72). Immune checkpoints are thus misused in Hodgkin lymphoma patients, via a process called "immunoediting" (73), causing T cell exhaustion. This leads to an immunosuppressive microenvironment and contributes to the survival of the tumor. Immunoediting involves three phases: 1) an initial elimination phase, which originally represents the concept of cancer immunosurveillance, eliminating the tumor by making use of both the innate and adaptive immune system. 2) The longest of three phases: an equilibrium phase between the host immune system and tumor cells that survived the elimination. The end result of this phase can be envisioned as the product of Darwinian selection: a population of tumor cells with reduced immunogenicity. 3) An escape phase in which tumor cell variants, that were selected out of the equilibrium phase, evade immune surveillance through a variety of immunoinhibitory mechanisms.

On a molecular level, overexpression of PD-L1 is partly caused by selective amplification of the 9p24.1 region in HRS cells, containing the PD-L1 gene. Moreover, the Jak2 locus is amplified causing Jak2 overexpression, which leads to subsequent transcriptional activation of PD-L1 (13,19,74). EBV infection leads to increased expression of PD-L1 gene and protein (19,75). Hodgkin lymphoma cell lines highly express a chromosomal fusion gene involving a CIITA MHC-II transcriptional activator, which leads to downregulation of surface HLA class II expression and overexpression of PD-L1 (19,76). Antigen presentation by MHC molecules is an important factor in the initiation of an immune response. HRS cells often lack HLA class I molecules, and therefore CD8+ T cells are not likely to be of much importance in immune checkpoint inhibition therapy in Hodgkin lymphoma. No association was found between MHC class I expression on HRS cells and PFS. However, MHC class II expression was found in 92% of complete responders to PD-1 blockade therapy. Furthermore, positive MHC class II expression on HRS cells in Hodgkin lymphoma patients was associated with elongated PFS when the anti-PD-1 antibody was received >12 months after autologous stem-cell transplantation (ASCT) (77). Altogether, HRS cells are able to evade the immune system by shutting down the immune response.

Results of anti-PD-1 antibodies pembrolizumab and nivolumab have been very promising for relapsed and refractory Hodgkin lymphoma patients. An 87% objective response rate (ORR) with a 20% complete response (CR) was achieved in refractory heavily pretreated Hodgkin lymphoma patients (19,78).

### PD-1-PD-L1 expression in solid tumors

The PD-1 blocking antibodies pembrolizumab and nivolumab have been approved by the US Food and Drug Administration (FDA) for patients with solid tumors including advanced unresectable melanoma and NSCLC (80). Predictive and progressive biomarkers are needed for the optimization of therapeutic benefit, minimization of risk of the autoimmune toxicities, and decrease of treatment costs (81). Predictive value can be determined by dividing the number of correctly predicted responders by the total number of patients with a positive biomarker result. Direct assessment of PD-L1 expression on tumor cells would be a logical biomarker for the prediction of durable efficacy to PD-1 blockade therapy in patients with solid tumors, like NSCLC and melanoma. However, soluble PD-1 or PD-L1 might be a better and less invasive manner to predict and/or determine the efficacy of immune checkpoint inhibitors on cancer therapy. Moreover, several other soluble factors, including IDO, CD83, and VEGF, have been described influencing the PD-1/PD-L1 pathway and might be of relevance as well.

#### sPD-L1

As mentioned before, sPD-L1 is often the product of proteolytic cleavage of the membrane-bound form of the molecule, or translation of alternative spliced mRNA. Cytokine treatment with IFN- $\alpha$ , IFN- $\gamma$ , or TNF- $\alpha$  is also positively associated with sPD-L1 secretion as well as membrane-bound PD-L1 expression in melanoma cells (1). Elevated sPD-L1 is associated with worse clinical outcome in many types of cancer (21–23,82). Studies about higher pretreatment levels of sPD-L1 also support this claim (1,83), however, rise in sPD-L1 after 5 months of treatment resulted in greater likelihood of partial responses after PD-1 inhibition (1). Upregulation of sPD-L1 after immune checkpoint inhibition may be caused by the changes in the TME as a result of the therapy-induced anti-tumor response, which causes enhanced production of cytokines including IFN- $\alpha$ , IFN- $\gamma$ , and TNF- $\alpha$  (1,84).

#### sPD-L2

In addition to sPD-L1, sPD-L2 is an important ligand for PD-1. Studies have suggested that sPD-L2 may play a role in the regulation of inflammation and autoimmunity. Costantini, et al. (85) described the predictive role of plasmatic biomarkers such as sPD-L2 in advanced NSCLC when patients were treated with nivolumab. sPD-L2 concentrations and variations showed no impact on clinical benefit, ORR, progression free survival (PFS) or overall survival (OS) in this study. However, it was found that lower concentrations of sPD-L2 at initial diagnosis and at nivolumab initiation were associated with patients presenting grade 3-4 toxicity. This means that sPD-L2 may be an effective biomarker in the prediction of immune-related toxicities rather than in the prediction of tumor response and clinical benefit (85)

#### sPD-1

In the development of several cancers, there is a strong interaction between sPD-1 and its ligand sPD-L1. Furthermore, these factors are present in elevated plasma levels in patients with a variety of cancers, including advanced pancreatic cancer (86), NSCLC (87), and pancreatic ductal adenocarcinoma (PDAC) (82). Assessment of the kinetics of sPD-1 before and after treatment with nivolumab shows the impact of treatment with anti-PD-1 antibodies on the production or destruction of sPD-1 and gives insight into clinical outcome. Treatment with nivolumab caused a decrease in sPD-1 levels of NSCLC patients after two cycles of treatment, which was also associated with a worse prognosis. This is in line with previous results of NSCLC patients with epidermal growth factor receptor (EGFR) mutations from Sorensen, Demuth, Weber, Sorensen, & Meldgaard (88).

#### IDO

In metastatic melanoma patients who were treated with anti-PD-1 agents, it was found that patients with the highest PD-1-PD-L1 interaction score and IDO-1-HLA-DR co-expression appeared to be more likely to respond to therapy and experienced significantly improved progression-free survival. This describes a connection between these factors (89). Furthermore, it was found that patients with advanced melanoma, treated with ipilimumab, an antibody that blocks CTLA-4, showed significant associations between baseline FoxP3 expression together with baseline IDO expression in tumor tissue, and clinical activity and increase in tumor-infiltrating lymphocytes (TILs) before and 3 weeks after start of treatment (p = 0.005). FoxP3 is a transcription factor, mainly expressed on CD4+ T cells, and has an immunosuppressive character because expression of Fox3P contributes to the suppression of many genes including IL-2 and effector T cell cytokines (90). The association between Fox3P together with baseline IDO expression and treatment with ipilimumab may be positive for clinical outcome (84,91). IDO expression in tumor tissue was assessed by IHC in tumor tissue, but in another study, in which IDO activity was assessed by serum kynurenine/tryptophan ratio, high IDO expression was associated with worse clinical outcome in NSCLC patients treated with nivolumab (92). This indicates that IDO activity may vary dependent on the assessment. (84). Previous studies have described co-expression of IDO and PD-L1 in several solid malignancies, including colon cancer and breast cancer (93–95). In a study on the relation between IDO and PD-L1 expression in cervical and vulvar squamous carcinomas (SCC), PD-L1 and IDO co-expression was observed in a subset of patients, which suggests a role for these factors in combination immunotherapy in cancer. The presence of invasion is also not expected to be of much benefit for combination treatment with anti-PD-L1 and anti-IDO agents (93).

#### CD83

In healthy human sera, sCD83 is present at low levels. However, patients with autoimmune disease or hematopoietic malignancies have elevated levels of sCD83. Recombinant soluble extracellular CD83 (rsCD83) mimics the natural sCD83, and binding of this molecule to the TLR4/MD-2 complex on monocytes induces production of anti-inflammatory mediators, such as IDO, leading to an immunosuppressive environment (96,97). Several solid tumors, including small cell lung cancer and other lung adenocarcinomas, have been shown to express CD83 (98). Moreover, polymorphisms of CD83, which were correlated with prognosis of cervical cancer, have been found in several cancers. Little is known about the relation between levels of sCD83 and prognosis in solid cancers (96,99).

#### VEGF

In a mouse model of colorectal cancer, it was found that the VEGF-A concentration in the TME in vivo was ~10 times higher compared to plasma levels of VEGF-A. After treatment targeting the VEGFA-VEGF receptor (VEGFR) axis with anti-VEGF-A antibodies, a significant reduction of PD-1 expression on

intratumoral CD8+ T cells was found and IFN-γ production in intratumoral CD8+ T cells was restored. Moreover, a combination of antiangiogenic agents such as anti-VEGF-A and immunotherapeutic approaches such as the use of anti-PD-1 induced a much stronger antitumor effect compared with therapy anti-VEGF-A alone or anti-PD-1 alone. Colorectal cancer patients showed overexpression of VEGF mRNA, which was associated with poorer survival than patients without overexpression. In a small cell lung cancer (SCLC) mouse model, mice were treated with a combination therapy of anti-PD-L1 and anti-VEGF agents, which restored the exhausted T cell phenotype of the mice and significantly improved PFS and OS. Thus, VEGF may serve as a predictive biomarker for the prognosis of several solid malignancies and anti-VEGF/anti-PD-L1 as well as anti-VEGFA/anti-PD-1 combination therapy synergistically improves clinical outcome in a SCLC mouse model and colorectal cancer patients, respectively (33,100,101).

# PD-1-PD-L1 expression in Hodgkin lymphoma

To our knowledge, no reports have been published about the potential role of sPD-L2 as a biomarker in Hodgkin lymphoma. However, sPD-L1, sPD-1, IDO, CD83, and VEGF have been described as important factors in the PD-1/PD-L1 pathway in Hodgkin lymphoma patients.

#### sPD-L1

Elevated mPD-L1 levels in classical Hodgkin lymphoma are associated with 9p24.1 amplification, which also leads to significantly shorter PFS for patients (102,103). Elevated sPD-L1 levels are often found in patients with several types of cancer compared to healthy individuals. This is also the case for Hodgkin lymphoma patients. Higher sPD-L1 levels in Hodgkin lymphoma patients are positively correlated with advanced stage and significantly shorter PFS, but not with OS. This suggests a potential role of sPD-L1 as an independent prognostic biomarker in classical Hodgkin lymphoma. Conflicting results have been obtained about the relation between EBV positivity and plasma sPD-L1 levels (103–105).

#### sPD-1

It is known that sPD-1 can prevent interaction between PD-1 and PD-L1 in vivo (106). In one study, Hodgkin lymphoma patients show high plasma levels of sPD-1 at diagnosis and before treatment with chemotherapy and radiotherapy, and these levels decrease as treatment proceeds. No correlation was found between sPD-1 and sPD-L1 expression. (107). To our knowledge, this is the only study to the role of sPD-1 as a potential biomarker in Hodgkin lymphoma.

#### IDO

In Hodgkin lymphoma patients, IDO is not expressed by HRS cells or lymphocytes. However, the molecule is active in the TME given its expression by macrophages, dendritic cells and vascular endothelial cells. Research supports that high expression of IDO is a significant prognostic predictor of bad clinical outcome in Hodgkin lymphoma. EBV and HIV-positive Hodgkin lymphoma patients show higher expression of IDO. High IDO expression is also associated with decreased CD4+ T cells and increased CD8+ T cells (108). Since CD8+ T cells recognize tumor cells through (neo)antigens presented by HLA class I molecules, this could be positive for clinical outcome. However, considering the fact that HLA class I molecules are often absent on HRS cells, CD8+ T cells are not likely to be of much importance in immune checkpoint inhibition therapy in

Hodgkin lymphoma. CD4+ T cells are more involved in the antitumor immune response in classical Hodgkin lymphoma, and interaction between PD-1 and CD4+ T cells is also enriched in the TME.

#### CD83

CD83 is expressed on HRS cells in lymph node biopsies of Hodgkin lymphoma patients. Recently, it was found that the membrane cleaved soluble form of CD83 is released into serum by Hodgkin lymphoma tumor cells, where the protein has an immune-inhibitory function by inhibiting T-cell proliferation. Decreases in circulating sCD83 were observed in patients who showed a CR after treatment with chemotherapy. Furthermore, research shows that CD4+ T cells that express CD83, expressed a higher level of PD-1 than CD83<sup>-</sup> T cells. This indicates that combination therapy of CD83 blockade and PD-1 blockade may enhance clinical response (30).

#### VEGF

VEGF can be produced by HRS cells, leading to upregulation of PD-L1 and PD-L2 expression levels (13,109). A combination of doxorubicin, bleomycin, vinblastine, and dacarbazine (ABVD) therapy in addition to anti-VEGF therapy may in fact be effective for Hodgkin lymphoma patients (109). However, elevated levels of VEGF, PD-L1, and PD-L2 are not significantly associated with event-free survival (EFS) as well as OS (109,110). Other studies suggest that high VEGF-A gene expression in classical Hodgkin lymphoma is associated with worse OS (111,112).

# Discussion

The aim of this study was to investigate the potential prognostic and predictive value of sPD-L1, sPD-L2, sPD-1, IDO, sCD83, and VEGF for PD-1 blockade therapy, and their impact on the PD-1/PL-L1 axis in Hodgkin lymphoma. Immune checkpoint therapy is shown to be very effective in Hodgkin lymphoma because the disease is characterized by a large TME, which forms an immunosuppressive, tumor tolerant environment with overexpression of PD-1 and PD-L1. Overexpressed PD-1 can be targeted by immunosuppressive agents such as nivolumab and pembrolizumab. Broader knowledge about the impact of these soluble factors on immune checkpoint inhibitors would make a significant contribution to treatment optimization. Soluble factors are preferred over the membrane-bound biomarkers because detection of biomarkers in plasma is easier and less invasive for the patient compared to a biopsy. The identification of potential responders and non-responders to immune checkpoint blockade therapy is very useful because like this, patients lose no time on ineffective therapy and the financial burden of unsuccessful treatment is limited (87,113). To this day, research has mainly been on the effect of the expression of soluble markers in solid malignancies and on tumor PD-L1 expression and current knowledge about the possible role of soluble biomarkers in the prediction and progression of Hodgkin lymphoma is limited (87).

In solid tumors, high pretreatment sPD-L1 levels are often associated with worse clinical outcome, but since rise in sPD-L1 levels during treatment have shown to contribute to better response to PD-1 blockade therapy. In Hodgkin lymphoma, elevated levels of sPD-L1 are associated with advanced stage and significantly shorter PFS. Therefore, we find sPD-L1 a valuable predictive biomarker for Hodgkin lymphoma. However, further investigation to pretreatment sPD-L1 levels and the kinetics of sPD-L1 in Hodgkin lymphoma during treatment is needed and might give insight into the exact role in the progression of the disease. In solid tumors, sPD-L2 has shown little predictive value for efficacy of immunotherapy, and no reports have been published about the potential role of sPD-L2 as a biomarker in Hodgkin lymphoma. sPD-1 expression is decreased in solid tumors after treatment with PD-1 inhibitors, which is also associated with a worse prognosis. Hodgkin lymphoma treatment with radiotherapy and chemotherapy decreases the elevated levels of sPD-1, but little is known about the predictive value for efficacy of PD-1 blockade therapy. However, current knowledge about sPD-1 in solid tumors is promising for sPD-1 as a predictive biomarker in Hodgkin lymphoma. The predictive value of IDO expression has been dichotomous in solid malignancies. However, in Hodgkin lymphoma, IDO expression is described as a significant prognostic predictor of bad clinical outcome. IDO is not expressed by HRS cells, but still has a large role in the TME. sCD83 expression in patients with solid malignancies had not been described in a predictive or progressive relation. However, in Hodgkin lymphoma, CR was associated with decreases in sCD83 levels after chemotherapy. VEGF overexpression in colorectal cancer patients was associated with poorer survival. In contrast, in Hodgkin lymphoma patients, no significant relation to VEGF expression and clinical outcome was found.

It is often observed that despite few impressive clinical successes, immunotherapy with a single-agent immune checkpoint inhibitor can not serve as a sufficiently effective therapy for many cancers (93,114,115). This might be the result of the complex composition of the TME, and the fact that other immunomodulatory molecules compensate for the loss of one immune checkpoint. The presence of the other factors is enough to continue the evasion of the immune surveillance by the tumor, even during PD-1/PD-L1 blockade therapy. Therefore, researchers have recently started to investigate the effects of

targeted combination immunotherapy for the treatment of solid malignancies (93,114). Promising effects have been observed in combination therapy blocking IDO or VEGF in combination with PD-1 or PD-L1 in solid malignancies. Therefore, these markers might also be of importance for combination immunotherapy in Hodgkin lymphoma. Little is known about sPD-L1, sPD-L2, sPD-1 and sCD83 in the field of combination therapy.

High expression of IDO in patients with solid malignancies has been associated with primary resistance to PD-1 blockade therapy. Therefore, combined IDO inhibition and PD-1 blockade may be very effective. IDO was also associated with decreased CD4+ cells and increased CD8+ cells in the TME of Hodgkin lymphoma (108), which might be an interesting aspect for combination immunotherapy in Hodgkin lymphoma. In solid tumors, a possible connection between IDO-1-HLA-DR co-expression and high PD-1-PD-L1 interaction was found and therefore this could be an interesting topic for future studies on the possible role of IDO in combination immunotherapy in Hodgkin lymphoma. Furthermore, baseline FoxP3 expression in combination with IDO expression might have a positive effect on clinical outcome in melanoma patients (84,91). Since Fox3P expression is also elevated in classical Hodgkin lymphoma patients (116), a combination therapy with anti-Fox3P agents and anti-IDO agents or anti-PD-L1 agents might have a positive effect on clinical outcome in Hodgkin lymphoma.

Combination immunotherapy with a combination anti-VEGF-A and anti-PD-1 has been very effective in solid malignancies (33,100,101). The effects of high VEGF expression in Hodgkin lymphoma patients are not yet clear but could be of importance for clinical outcome and as a target immunotherapy. Given the finding that CD4+ T cells that express CD83, express a higher level of PD-1 than CD83<sup>-</sup> T cells, a combination of CD83 blockade and PD-1 blockade therapy might be also of clinical value in Hodgkin lymphoma patients.

After assessment with immunohistochemistry, the PD-1 or PD-L1 axis is not always accurately reflected, which could be partly caused by the poor uniformity in the PD-L1 immunohistochemistry anti bodies (14,80). A variety of tumors, including melanoma, renal cell carcinoma, NSCLC, and bladder cancer are associated with good clinical responses to immune checkpoint inhibition with anti-PD-1 agents. However, PD-L1 as a predictive biomarker for these cancers results in a wide range of PD-L1 IHC expression (14-100%), which highlights the issues with PD-L1 as a valuable, predictive biomarker for anti-PD-1 therapy (14,117–119).

Further studies aiming to identify predictive and progressive biomarkers for treatment of Hodgkin lymphoma with immune checkpoint inhibitors should include the kinetics of sPD-L1 in Hodgkin lymphoma during treatment, the effects of EBV and HIV infection on the TME and the role of sPD-L2 in the progression of Hodgkin lymphoma.

Based on our findings, we believe that sPD-L1 and sPD-1 are promising independent predictive biomarkers for Hodgkin lymphoma. Furthermore, combination therapy of anti-IDO with anti-PD-1 agents as well as anti-VEGF with anti-PD-1 antibodies might provide positive clinical outcome for Hodgkin lymphoma patients.

# Conflict of interest

There was no conflict of interest.

# Acknowledgements

In this section I would like to take the chance to thank certain people who helped me in the process of this thesis. Firstly, dr Lydia Visser for guiding and supporting me through the learning process that came with the writing of this thesis. Secondly, Johanna Veldman for the great feedback and guidance. Thirdly, prof. Dr. J.H.M. van den Berg for being my first supervisor.

### References

- 1. Zhou J, Mahoney KM, Giobbie-Hurder A, Zhao F, Lee S, Liao X, et al. Soluble PD-L1 as a biomarker in malignant melanoma treated with checkpoint blockade. Cancer Immunol Res. 2017;5(6):480–92.
- 2. Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and Its Ligands in Tolerance and Immunity. Annu Rev Immunol. 2008;26(1):677–704.
- 3. Zhu X, Lang J. Soluble PD-1 and PD-L1: Predictive and prognostic significance in cancer. Oncotarget. 2017;8(57):97671–82.
- 4. Topalian SL, Taube JM, Anders RA PD. Mechanism-driven biomarkers to guide immune checkpoint blockade in cancer therapy. Nat Rev Cancer. 2017;176(1):139–48.
- 5. Pardoll DM. Cancer and the Immune System: Basic Concepts and Targets for Intervention. Semin Oncol. 2015;42(4):523–38.
- 6. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer. 2012;42(4):587–600.
- Hamanishi J, Mandai M, Iwasaki M, Okazaki T, Tanaka Y, Yamaguchi K, et al. Programmed cell death 1 ligand 1 and tumor-infiltrating CD8+ T lymphocytes are prognostic factors of human ovarian cancer. Proc Natl Acad Sci U S A. 2007;104(9):3360–5.
- Ghebeh H, Tulbah A, Mohammed S, ElKum N, Bin Amer SM, Al-Tweigeri T, et al. Expression of B7-H1 in breast cancer patients is strongly associated with high proliferative Ki-67-expressing tumor cells. Int J Cancer. 2007;121(4):751–8.
- 9. Thanarajasingam G, Thanarajasingam U, Ansell SM. Immune checkpoint blockade in lymphoid malignancies. FEBS J. 2016;283(12):2233–44.
- 10. Wei SC, Duffy CR, Allison JP. Fundamental mechanisms of immune checkpoint blockade therapy. Cancer Discov. 2018;8(9):1069–86.
- 11. Chen BJ, Bjoern Chapuy, Jing Ouyang, Heather H Sun, Margaretha GM Roemer, Mina L Xu, Hongbo Yu, Christopher DM Fletcher, FRCPath1, Gordon J. Freeman, Margaret A Shipp and SJR. PD-L1 Expression is Characteristic of a subset of. Clin Cancer Res. 2014;19(13):3462–73.
- 12. Hino R, Kabashima K, Kato Y, Yagi H, Nakamura M, Honjo T, et al. Tumor cell expression of programmed cell death-1 ligand 1 is a prognostic factor for malignant melanoma. Cancer. 2010;116(7):1757–66.
- Veldman J, Visser L, Berg A van den, Diepstra A. Primary and acquired resistance mechanisms to immune checkpoint inhibition in Hodgkin lymphoma. Cancer Treat Rev [Internet]. 2020;82(November 2019):101931. Available from: https://doi.org/10.1016/j.ctrv.2019.101931
- Patel SP, Kurzrock R. PD-L1 expression as a predictive biomarker in cancer immunotherapy [Internet]. Vol. 14, Molecular Cancer Therapeutics. American Association for Cancer Research Inc.; 2015 [cited 2020 Mar 25]. p. 847– 56. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25695955
- 15. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Mutational landscape determines sensitivity to PD-1 blockade in non–small cell lung cancer. Science (80-). 2016;348(6230):124–8.
- 16. DT L, Durham J, Smith K, Wang H, Bartlett B, Aulakh L, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science (80-). 2017;357(6349):409–13.
- 17. Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H, Eyring AD, et al. PD-1 blockade in tumors with mismatch-repair deficiency. N Engl J Med. 2015;372(26):2509–20.
- 18. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. Vol. 144, Cell. 2011. p. 646–74.
- 19. Lin RJ, Diefenbach CS. Checkpoint Inhibition in Hodgkin Lymphoma: Saving the Best for Last? Oncology (Williston Park). 2016;30(10):914–20.
- Chen Y, Wang Q, Shi B, Xu P, Hu Z, Bai L, et al. Development of a sandwich ELISA for evaluating soluble PD-L1 (CD274) in human sera of different ages as well as supernatants of PD-L1 + cell lines. Cytokine. 2011 Nov;56(2):231–8.
- 21. Frigola X, Inman BA, Lohse CM, Krco CJ, Cheville JC, Thompson RH, et al. Identification of a soluble form of B7-H1

that retains immunosuppressive activity and is associated with aggressive renal cell carcinoma. Clin Cancer Res [Internet]. 2011 Apr 1 [cited 2020 Mar 27];17(7):1915–23. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21355078

- 22. Rossille D, Gressier M, Damotte D, Maucort-Boulch D, Pangault C, Semana G, et al. High level of soluble programmed cell death ligand 1 in blood impacts overall survival in aggressive diffuse large B-cell lymphoma: Results from a French multicenter clinical trial. Leukemia. 2014 Dec 11;28(12):2367–75.
- 23. Finkelmeier F, Canli Ö, Tal A, Pleli T, Trojan J, Schmidt M, et al. High levels of the soluble programmed death-ligand (sPD-L1) identify hepatocellular carcinoma patients with a poor prognosis. Eur J Cancer [Internet]. 2016 May 1 [cited 2020 Mar 27];59:152–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27039170
- 24. Okuma Y, Hosomi Y, Nakahara Y, Watanabe K, Sagawa Y, Homma S. High plasma levels of soluble programmed cell death ligand 1 are prognostic for reduced survival in advanced lung cancer. Lung Cancer [Internet]. 2017 Feb 1 [cited 2020 Mar 27];104:1–6. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28212990
- 25. Zheng Z, Bu Z, Liu X, Zhang L, Li Z, Wu A, et al. Level of circulating PD-L1 expression in patients with advanced gastric cancer and its clinical implications. Chinese J Cancer Res. 2014 Feb 1;26(1):104–11.
- 26. Sunshine J, Taube JM. PD-1/PD-L1 inhibitors. Curr Opin Pharmacol. 2015;23:32–8.
- Lipson EJ, Forde PM, Hammers HJ, Emens LA, Taube JM, Topalian SL. Antagonists of PD-1 and PD-L1 in Cancer Treatment. Semin Oncol [Internet]. 2015;42(4):587–600. Available from: http://dx.doi.org/10.1053/j.seminoncol.2015.05.013
- 28. Fallarino F, Grohmann U, Vacca C, Orabona C, Spreca A, Fioretti MC, et al. T cell apoptosis by kynurenines. In: Advances in Experimental Medicine and Biology. 2003. p. 183–90.
- 29. Fallarino F, Grohmann U, You S, McGrath BC, Cavener DR, Vacca C, et al. Tryptophan catabolism generates autoimmune-preventive regulatory T cells. Transpl Immunol. 2006 Dec 1;17(1):58–60.
- 30. Li Z, Ju X, Lee K, Clarke C, Hsu JL, Abadir E, et al. Cd83 is a new potential biomarker and therapeutic target for hodgkin lymphoma. Haematologica. 2018;103(4):655–65.
- 31. Hock BD, Kato M, McKenzie JL, Hart DNJ. A soluble form of CD83 is released from activated dendritic cells and B lymphocytes, and is detectable in normal human sera. Int Immunol. 2001;13(7):959–67.
- 32. Gabrilovich DI, Chen HL, Girgis KR, Cunningham HT, Meny GM, Nadaf S, et al. Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. Nat Med. 1996 Oct;2(10):1096–103.
- 33. Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet AL, et al. VEGF-A modulates expression of inhibitory checkpoints on CD8++ T cells in tumors. J Exp Med. 2015;212(2):139–48.
- American, Cancer, Society. Hodgkin lymphoma Statistics | American Cancer Society Cancer Facts & Cancer & Can
- 35. Wein F, Küppers R. The role of T cells in the microenvironment of Hodgkin lymphoma. J Leukoc Biol. 2016 Jan;99(1):45–50.
- 36. Steidl C, Connors JM, Gascoyne RD. Molecular pathogenesis of hodgkin's lymphoma: Increasing evidence of the importance of the microenvironment. J Clin Oncol. 2011;29(14):1812–26.
- 37. Weiss LM, Strickler JG, Warnke RA, Purtilo DT, Sklar J. Epstein-Barr viral DNA in tissues of Hodgkin's disease. Am J Pathol. 1987;129(1):86–91.
- 38. Khan G. Epstein-Barr virus, cytokines, and inflammation: A cocktail for the pathogenesis of Hodgkin's lymphoma? Vol. 34, Experimental Hematology. 2006. p. 399–406.
- 39. Ansell SM. Hodgkin lymphoma: 2016 update on diagnosis, risk-stratification, and management. Am J Hematol. 2016;91(4):434–42.
- 40. Alexander FE, Jarrett RF, Lawrence D, Armstrong AA, Freeland J, Gokhale DA, et al. Risk factors for Hodgkin's disease by Epstein-Barr virus (EBV) status: Prior infection by EBV and other agents. Br J Cancer. 2000;82(5):1117–21.
- 41. Campo E, Swerdlow SH, Harris NL, Pileri S, Stein H, Jaffe ES. The 2008 WHO classification of lymphoid neoplasms and beyond: evolving concepts and practical applications. Blood. 2011;117(19):5019–32.

- 42. Marafioti T, Hummel M, Foss HD, Laumen H, Korbjuhn P, Anagnostopoulos I, et al. Hodgkin and Reed-Sternberg cells represent an expansion of a single clone originating from a germinal center B-cell with functional immunoglobulin gene rearrangements but defective immunoglobulin transcription. Blood. 2000 Feb 15;95(4):1443–50.
- 43. Falini B, Stein H, Pileri S, Canino S, Farabbi R, MARTELLI MF, et al. Expression of lymphoid-associated antigens on Hodgkin's and Reed-Sternberg cells of Hodgkin's disease. An immunocytochemical study on lymph node cytospins using monoclonal antibodies. Histopathology [Internet]. 1987 Dec 1 [cited 2020 Mar 22];11(12):1229–42. Available from: http://doi.wiley.com/10.1111/j.1365-2559.1987.tb01869.x
- 44. Schwering I, Bräuninger A, Klein U, Jungnickel B, Tinguely M, Diehl V, et al. Loss of the B-lineage-specific gene expression program in Hodgkin and Reed-Sternberg cells of Hodgkin lymphoma. Blood. 2003 Feb 15;101(4):1505–12.
- 45. Deutsch YE, Tadmor T, Podack ER, Rosenblatt JD. CD30: an important new target in hematologic malignancies. Leuk Lymphoma [Internet]. 2011 Sep 27 [cited 2020 Mar 22];52(9):1641–54. Available from: http://www.tandfonline.com/doi/full/10.3109/10428194.2011.574761
- 46. Pileri SA, Ascani S, Leoncini L, Sabattini E, Zinzani PL, Piccaluga PP, et al. Hodgkin's lymphoma: The pathologist's viewpoint. Vol. 55, Journal of Clinical Pathology. BMJ Publishing Group; 2002. p. 162–76.
- 47. Poppema S, Bhan AK, Reinherz EL, Posner MR, Schlossman SF. In situ immunologic characterization of cellular constituents in lymph nodes and spleens involved by Hodgkin's disease. Blood. 1982;59(2):226–32.
- Poppema S, Lai R, Visser L, Yan XJ. CD45 (leucocyte common antigen) expression in T and B lymphocyte subsets. Leuk Lymphoma [Internet]. 1996 Jan [cited 2020 Mar 23];20(3–4):217–22. Available from: http://www.ncbi.nlm.nih.gov/pubmed/8624459
- Ma Y, Visser L, Blokzijl T, Harms G, Atayar Ç, Poppema S, et al. The CD4+CD26- T-cell population in classical Hodgkin's lymphoma displays a distinctive regulatory T-cell profile. Lab Investig [Internet]. 2008 May [cited 2020 Mar 23];88(5):482–90. Available from: http://www.ncbi.nlm.nih.gov/pubmed/18362907
- 50. Van Den Berg A, Visser L, Poppema S. High expression of the CC chemokine TARC in Reed-Sternberg cells: A possible explanation for the characteristic T-cell infiltrate in Hodgkin's lymphoma. Am J Pathol [Internet]. 1999 Jun [cited 2020 Mar 23];154(6):1685–91. Available from: http://www.ncbi.nlm.nih.gov/pubmed/10362793
- 51. Buri C, Körner M, Schärli P, Cefai D, Uguccioni M, Mueller C, et al. CC chemokines and the receptors CCR3 and CCR5 are differentially expressed in the nonneoplastic leukocytic infiltrates of Hodgkin disease. Blood [Internet]. 2001 Mar 15 [cited 2020 Mar 23];97(6):1543–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/11238088
- 52. Ohshima K, Tutiya T, Yamaguchi T, Suzuki K, Suzumiya J, Kawasaki C, et al. Infiltration of Th1 and Th2 lymphocytes around Hodgkin and Reed-Sternberg (H&RS) cells in Hodgkin disease: Relation with expression of CXC and CC chemokines on H&RS cells. Int J Cancer [Internet]. 2002 Apr 1 [cited 2020 Mar 23];98(4):567–72. Available from: http://doi.wiley.com/10.1002/ijc.10218
- 53. Peh SC, Kim LH, Poppema S. TARC, a CC chemokine, is frequently expressed in classic Hodgkin's lymphoma but not in NLP Hodgkin's lymphoma, T-cell-rich B-cell lymphoma, and most cases of anaplastic large cell lymphoma. Am J Surg Pathol. 2001;25(7):925–9.
- 54. Hedvat C V., Jaffe ES, Qin J, Filippa DA, Cordon-Cardo C, Tosato G, et al. Macrophage-derived chemokine expression in classical Hodgkin's lymphoma: Application of tissue microarrays. Mod Pathol [Internet]. 2001 Dec [cited 2020 Mar 23];14(12):1270–6. Available from: http://www.ncbi.nlm.nih.gov/pubmed/11743050
- 55. Fischer M, Juremalm M, Olsson N, Backlin C, Sundström C, Nilsson K, et al. Expression of CCL5/RANTES by Hodgkin and reed-sternberg cells and its possible role in the recruitment of mast cells into lymphomatous tissue. Int J Cancer [Internet]. 2003 Nov 1 [cited 2020 Mar 23];107(2):197–201. Available from: http://www.ncbi.nlm.nih.gov/pubmed/12949794
- 56. Gerdes J, Kretschmer C, Zahn G, Ernst M, Jones DB, Flad HD. Immunoenzymatic assessment of interferon-γ in hodgkin and sternberg-reed cells. Cytokine. 1990;2(4):307–10.
- 57. Teruya-Feldstein J, Jaffe ES, Burd PR, Kingma DW, Setsuda JE, Tosato G. Differential chemokine expression in tissues involved by Hodgkin's disease: direct correlation of eotaxin expression and tissue eosinophilia. Blood [Internet]. 1999 Apr 15 [cited 2020 Mar 23];93(8):2463–70. Available from: http://www.ncbi.nlm.nih.gov/pubmed/10194423
- 58. Ma Y, Visser L, Roelofsen H, De Vries M, Diepstra A, Van Imhoff G, et al. Proteomics analysis of Hodgkin lymphoma: Identification of new players involved in the cross-talk between HRS cells and infiltrating lymphocytes. Blood

[Internet]. 2008 Feb 15 [cited 2020 Mar 23];111(4):2339–46. Available from: http://www.ncbi.nlm.nih.gov/pubmed/18070985

- 59. Hsu PL, Lin YC, Hsu SM. Expression of macrophage colony-stimulating factor (M-CSF) in two Hodgkin's Reed-Sternberg (H-RS) cell lines, HDLM-1 and KM-H2, and in H-RS cells in tissues. Int J Hematol [Internet]. 1991 Aug [cited 2020 Mar 23];54(4):315–26. Available from: http://www.ncbi.nlm.nih.gov/pubmed/1838015
- 60. Truman LA, Ford CA, Pasikowska M, Pound JD, Wilkinson SJ, Dumitriu IE, et al. CX3CL 1/fractalkine is released from apoptotic lymphocytes to stimulate macrophage chemotaxis. Blood. 2008 Dec 15;112(13):5026–36.
- 61. Skinnider BF, Mak TW. The role of cytokines in classical Hodgkin lymphoma [Internet]. Vol. 99, Blood. 2002 [cited 2020 Mar 23]. p. 4283–97. Available from: http://www.ncbi.nlm.nih.gov/pubmed/12036854
- 62. Jundt F, Anagnostopoulos I, Bommert K, Emmerich F, Müller G, Foss HD, et al. Hodgkin/Reed-Sternberg cells induce fibroblasts to secrete eotaxin, a potent chemoattractant for T cells and eosinophils. Blood. 1999 Sep 15;94(6):2065–71.
- 63. De Bruin ML, Sparidans J, Van't Veer MB, Noordijk EM, Louwman MWJ, Zijlstra JM, et al. Breast cancer risk in female survivors of Hodgkin's lymphoma: Lower risk after smaller radiation volumes. J Clin Oncol [Internet]. 2009 Sep 10 [cited 2020 Mar 23];27(26):4239–46. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19667275
- 64. Koontz MZ, Horning SJ, Balise R, Greenberg PL, Rosenberg SA, Hoppe RT, et al. Risk of therapy-related secondary leukemia in hodgkin lymphoma: The stanford university experience over three generations of clinical trials. J Clin Oncol [Internet]. 2013 Feb 10 [cited 2020 Mar 23];31(5):592–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/23295809
- 65. Broadfoot J, Johnson PWM. Response-adapted therapy in Hodgkin lymphoma. Hematol Oncol [Internet]. 2017 Jun 1 [cited 2020 Mar 23];35:33–6. Available from: http://doi.wiley.com/10.1002/hon.2398
- Van Nimwegen FA, Schaapveld M, Cutter DJ, Janus CPM, Krol ADG, Hauptmann M, et al. Radiation dose-response relationship for risk of coronary heart disease in survivors of Hodgkin lymphoma. J Clin Oncol [Internet]. 2016 Jan 20 [cited 2020 Mar 23];34(3):235–43. Available from: http://www.ncbi.nlm.nih.gov/pubmed/26573075
- 67. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SAJR, Behjati S, Biankin A V., et al. Signatures of mutational processes in human cancer. Nature. 2013;500(7463):415–21.
- 68. Jezeršek Novaković B. Checkpoint inhibitors in Hodgkin's lymphoma. Eur J Haematol [Internet]. 2016 Apr 1 [cited 2020 Mar 23];96(4):335–43. Available from: http://doi.wiley.com/10.1111/ejh.12697
- 69. Villasboas JC, Ansell S. Checkpoint inhibition: Programmed cell death 1 and programmed cell death 1 ligand inhibitors in Hodgkin lymphoma [Internet]. Vol. 22, Cancer Journal (United States). Lippincott Williams and Wilkins; 2016 [cited 2020 Mar 23]. p. 17–22. Available from: http://www.ncbi.nlm.nih.gov/pubmed/26841012
- Hawkes EA, Grigg A, Chong G. Programmed cell death-1 inhibition in lymphoma [Internet]. Vol. 16, The Lancet Oncology. Lancet Publishing Group; 2015 [cited 2020 Mar 23]. p. e234–45. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25943068
- 71. Ioachim HL, Cooper MC, Hellman GC. Hodgkin's disease and the acquired immunodeficiency syndrome. Vol. 101, Annals of Internal Medicine. American College of Physicians; 1984. p. 876–7.
- 72. Yamamoto R, Nishikori M, Kitawaki T, Sakai T, Hishizawa M, Tashima M, et al. PD-1 PD-1 ligand interaction contributes to immunosuppressive microenvironment of Hodgkin lymphoma. Blood [Internet]. 2008 Mar 15 [cited 2020 Mar 24];111(6):3220–4. Available from: http://www.ncbi.nlm.nih.gov/pubmed/18203952
- Dunn GP, Old LJ, Schreiber RD. The Three Es of Cancer Immunoediting. Annu Rev Immunol [Internet]. 2004 Apr 19 [cited 2020 Mar 26];22(1):329–60. Available from: http://www.annualreviews.org/doi/10.1146/annurev.immunol.22.012703.104803
- 74. Green MR, Monti S, Rodig SJ, Juszczynski P, Currie T, O'Donnell E, et al. Integrative analysis reveals selective 9p24.1 amplification, increased PD-1 ligand expression, and further induction via JAK2 in nodular sclerosing Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. Blood [Internet]. 2010 Oct 28 [cited 2020 Mar 24];116(17):3268–77. Available from: http://www.ncbi.nlm.nih.gov/pubmed/20628145
- 75. Green MR, Rodig S, Juszczynski P, Ouyang J, Sinha P, O'Donnell E, et al. Constitutive AP-1 activity and EBV infection induce PD-l1 in Hodgkin lymphomas and posttransplant lymphoproliferative disorders: Implications for targeted therapy. Clin Cancer Res [Internet]. 2012 Mar 15 [cited 2020 Mar 24];18(6):1611–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/22271878

- 76. Steidl C, Shah SP, Woolcock BW, Rui L, Kawahara M, Farinha P, et al. MHC class II transactivator CIITA is a recurrent gene fusion partner in lymphoid cancers. Nature [Internet]. 2011 Mar 17 [cited 2020 Mar 24];471(7338):377–83. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21368758
- 77. Roemer MGM, Redd RA, Cader Z, Pak CJ, Abdelrahman S, Ouyang J, et al. Major Histocompatibility Complex Class II and Programmed Death Ligand 1 Expression Predict Outcome After Programmed Death 1 Blockade in Classic Hodgkin Lymphoma. J Clin Oncol [Internet]. 2018 [cited 2020 Apr 2];36(10):942–50. Available from: https://doi.org/10.1200/JCO.2017.
- 78. Ansell SM, Lesokhin AM, Borrello I, Halwani A, Scott EC, Gutierrez M, et al. PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma. N Engl J Med [Internet]. 2015 Jan 22 [cited 2020 Apr 3];372(4):311–9. Available from: http://www.nejm.org/doi/10.1056/NEJMoa1411087
- 79. Goodman A, Patel SP, Kurzrock R. PD-1–PD-L1 immune-checkpoint blockade in B-cell lymphomas. 2017 [cited 2020 Apr 3]; Available from: www.nature.com/nrclinonc
- 80. Gibney GT, Weiner LM, Atkins MB. Predictive biomarkers for checkpoint inhibitor-based immunotherapy
  [Internet]. Vol. 17, The Lancet Oncology. Lancet Publishing Group; 2016 [cited 2020 Mar 25]. p. e542–51. Available
  from: http://www.ncbi.nlm.nih.gov/pubmed/27924752
- 81. Hamid O, Robert C, Daud A, Hodi FS, Hwu W-J, Kefford R, et al. Safety and Tumor Responses with Lambrolizumab (Anti–PD-1) in Melanoma. N Engl J Med [Internet]. 2013 Jul 11 [cited 2020 Mar 25];369(2):134–44. Available from: http://www.nejm.org/doi/10.1056/NEJMoa1305133
- 82. Bian B, Fanale D, Dusetti N, Roque J, Pastor S, Chretien AS, et al. Prognostic significance of circulating PD-1, PD-L1, pan-BTN3As, BTN3A1 and BTLA in patients with pancreatic adenocarcinoma. Oncoimmunology. 2019 Apr 3;8(4).
- Okuma Y, Wakui H, Utsumi H, Sagawa Y, Hosomi Y, Kuwano K, et al. Soluble Programmed Cell Death Ligand 1 as a Novel Biomarker for Nivolumab Therapy for Non–Small-cell Lung Cancer. Clin Lung Cancer [Internet]. 2018 Sep 1 [cited 2020 Mar 28];19(5):410-417.e1. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29859759
- 84. Nakamura Y. Biomarkers for Immune Checkpoint Inhibitor-Mediated Tumor Response and Adverse Events. Front Med. 2019;6(119).
- 85. Costantini A, Julie C, Dumenil C, Elias-Rodzewicz ZH, Tisserand J, Dumoulin J, et al. Predictive role of plasmatic biomarkers in advanced non-small cell lung cancer treated by nivolumab. 2018 [cited 2020 Mar 30]; Available from: https://doi.org/10.1080/2162402X.2018.1452581
- Kruger S, Legenstein ML, Rösgen V, Haas M, Modest DP, Westphalen CB, et al. Serum levels of soluble programmed death protein 1 (sPD-1) and soluble programmed death ligand 1 (sPD-L1) in advanced pancreatic cancer.
  Oncoimmunology [Internet]. 2017 May 4 [cited 2020 Apr 1];6(5):e1310358. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28638732
- Meyo MT, Jouinot A, Giroux-Leprieur E, Fabre E, Wislez M, Alifano M, et al. Predictive Value of Soluble PD-1, PD-L1, VEGFA, CD40 Ligand and CD44 for Nivolumab Therapy in Advanced Non-Small Cell Lung Cancer: A Case-Control Study. Cancers (Basel) [Internet]. 2020 [cited 2020 Apr 1];12:473. Available from: www.mdpi.com/journal/cancers
- 88. Sorensen SF, Demuth C, Weber B, Sorensen BS, Meldgaard P. Increase in soluble PD-1 is associated with prolonged survival in patients with advanced EGFR-mutated non-small cell lung cancer treated with erlotinib. Lung Cancer [Internet]. 2016;100:77–84. Available from: http://dx.doi.org/10.1016/j.lungcan.2016.08.001
- 89. Johnson DB, Bordeaux J, Kim JY, Vaupel C, Rimm DL, Ho TH, et al. Quantitative spatial profiling of PD-1/PD-L1 interaction and HLA-DR/IDO-1 predicts improved outcomes of anti-PD-1 therapies in metastatic melanoma. Clin Cancer Res [Internet]. 2018 Nov 1 [cited 2020 Mar 31];24(21):5250–60. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30021908
- 90. Kim CH. FOXP3 and its role in the immune system. Adv Exp Med Biol. 2009;665:17–29.
- 91. Hamid O, Schmidt H, Nissan A, Ridolfi L, Aamdal S, Hansson J, et al. A prospective phase II trial exploring the association between tumor microenvironment biomarkers and clinical activity of ipilimumab in advanced melanoma [Internet]. Vol. 9, Journal of Translational Medicine. 2011 [cited 2020 Mar 28]. Available from: http://www.translational-medicine.com/content/9/1/204
- 92. Botticelli A, Cerbelli B, Lionetto L, Zizzari I, Salati M, Pisano A, et al. Can IDO activity predict primary resistance to anti-PD-1 treatment in NSCLC? J Transl Med [Internet]. 2018 Aug 6 [cited 2020 Mar 28];16(1):219. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30081936
- 93. Chinn Z, Stoler MH, Mills AM. PD-L1 and IDO expression in cervical and vulvar invasive and intraepithelial

squamous neoplasias: implications for combination immunotherapy. Histopathology [Internet]. 2019 Jan 1 [cited 2020 Apr 3];74(2):256–68. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30067880

- 94. Dill EA, Dillon PM, Bullock TN, Mills AM. IDO expression in breast cancer: an assessment of 281 primary and metastatic cases with comparison to PD-L1. Mod Pathol [Internet]. 2018 Oct 1 [cited 2020 Apr 3];31(10):1513–22. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29802358
- 95. Friedman K, Brodsky AS, Lu S, Wood S, Gill AJ, Lombardo K, et al. Medullary carcinoma of the colon: A distinct morphology reveals a distinctive immunoregulatory microenvironment. Mod Pathol. 2016 May 1;29(5):528–41.
- 96. Caminschi I, Shortman K, Hall E, Kroczek RA, Li Z, Ju X, et al. CD83: Activation Marker for Antigen Presenting Cells and Its Therapeutic Potential. Front Immunol [Internet]. 2019 [cited 2020 Mar 31];10(1312). Available from: www.frontiersin.org
- 97. Horvatinovich JM, Grogan EW, Norris M, Steinkasserer A, Lemos H, Mellor AL, et al. Soluble CD83 Inhibits T Cell Activation by Binding to the TLR4/MD-2 Complex on CD14 + Monocytes . J Immunol [Internet]. 2017 Mar 15 [cited 2020 Mar 31];198(6):2286–301. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28193829
- 98. Baleeiro RB, Bergami-Santos PC, Tomiyoshi MY, Gross JL, Haddad F, Pinto CAL, et al. Expression of a dendritic cell maturation marker CD83 on tumor cells from lung cancer patients and several human tumor cell lines: Is there a biological meaning behind it? Cancer Immunol Immunother. 2008 Feb 13;57(2):265–70.
- Yu KJ, Rader JS, Borecki I, Zhang Z, Hildesheim A. CD83 polymorphisms and cervical cancer risk. Gynecol Oncol [Internet]. 2009 Aug [cited 2020 Mar 31];114(2):319–22. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19446866
- Ishigami S-1, Arii S, Furutani M, Niwano M, Harada T, Mizumoto M, et al. Predictive value of vascular endothelial growth factor (VEGF) in metastasis and prognosis of human colorectal cancer. Vol. 78410, Britsh Jourmal of Cancer. 1998.
- Meder L, Schuldt P, Thelen M, Schmitt A, Dietlein F, Klein S, et al. Combined VEGF and PD-L1 blockade displays synergistic treatment effects in an autochthonous mouse model of small cell lung cancer. Cancer Res [Internet].
   2018 [cited 2020 Apr 3];78(15):4270–81. Available from: http://cancerres.aacrjournals.org/
- 102. Roemer MGM, Advani RH, Ligon AH, Natkunam Y, Redd RA, Homer H, et al. PD-L1 and PD-L2 genetic alterations define classical hodgkin lymphoma and predict outcome. J Clin Oncol [Internet]. 2016 Aug 10 [cited 2020 Apr 1];34(23):2690–7. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27069084
- 103. Guo X, Wang J, Jin J, Chen H, Zhen Z, Jiang W, et al. High Serum Level of Soluble Programmed Death Ligand 1 is Associated With a Poor Prognosis in Hodgkin Lymphoma. Transl Oncol [Internet]. 2018 [cited 2020 Apr 1];11:779– 85. Available from: https://doi.org/10.1016/j.tranon.2018.03.012
- 104. Paydas S, Bağır E, Seydaoglu G, Ercolak V, Ergin M. Programmed death-1 (PD-1), programmed death-ligand 1 (PD-L1), and EBV-encoded RNA (EBER) expression in Hodgkin lymphoma. Ann Hematol. 2015 Sep 6;94(9):1545–52.
- 105. Hohaus S, Santangelo R, Giachelia M, Vannata B, Massini G, Cuccaro A, et al. The viral load of Epstein-Barr virus (EBV) DNA in peripheral blood predicts for biological and clinical characteristics in Hodgkin lymphoma. Clin Cancer Res [Internet]. 2011 May 1 [cited 2020 Apr 1];17(9):2885–92. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21478335
- 106. Gu D, Ao X, Yang Y, Chen Z, Xu X. Soluble immune checkpoints in cancer: Production, function and biological significance. Vol. 6, Journal for ImmunoTherapy of Cancer. BioMed Central Ltd.; 2018. p. 1–14.
- 107. da Silva PB, Real JM, Ferreira LRP, Esteves GH, Brito F do N, Baiocchi OCG. Soluble PD-1 and PD-L1 as potential biomarkers for classical Hodgkin lymphoma. Vol. 36, Hematological Oncology. John Wiley and Sons Ltd; 2018. p. 709–12.
- 108. Choe J-Y, Yun JY, Kyoung Jeon Y, Kim H, Park G, Huh JR, et al. Indoleamine 2,3-dioxygenase (IDO) is frequently expressed in stromal cells of Hodgkin lymphoma and is associated with adverse clinical features: a retrospective cohort study [Internet]. Vol. 14. 2014 [cited 2020 Mar 31]. Available from: http://www.biomedcentral.com/1471-2407/14/335
- 109. Koh YW, Han JH, Yoon DH, Suh C, Huh J. PD-L1 expression correlates with VEGF and microvessel density in patients with uniformly treated classical Hodgkin lymphoma. Ann Hematol [Internet]. 2017 Nov 1 [cited 2020 Mar 31];96(11):1883–90. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28842748
- 110. Dimtsas GS, Georgiadi EC, Karakitsos P, Vassilakopoulos TP, Thymara I, Korkolopoulou P, et al. Prognostic significance of immunohistochemical expression of the angiogenic molecules vascular endothelial growth factor-A,

vascular endothelial growth factor receptor-1 and vascular endothelial growth factor receptor-2 in patients with classical Hodgkin lymphoma. Leuk Lymphoma. 2014 Mar;55(3):558–64.

- 111. Linke F, Harenberg M, Zaunig S, Von Bonin F, Arlt A, Szczepanowski M, et al. Microenvironmental interactions between endothelial and lymphoma cells: a role for the canonical WNT pathway in Hodgkin lymphoma. Leukemia [Internet]. 2017 [cited 2020 Mar 31];31:361–72. Available from: www.nature.com/leu
- 112. Steidl C, Lee T, Shah SP, Farinha P, Han G, Nayar T, et al. Tumor-associated macrophages and survival in classic Hodgkin's lymphoma. N Engl J Med [Internet]. 2010 Mar 11 [cited 2020 Mar 31];362(10):875–85. Available from: http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=2897174&tool=pmcentrez&rendertype=abstract
- 113. Tartari F, Santoni M, Burattini L, Mazzanti P, Onofri A, Berardi R. Economic sustainability of anti-PD-1 agents nivolumab and pembrolizumab in cancer patients: Recent insights and future challenges. Vol. 48, Cancer Treatment Reviews. W.B. Saunders Ltd; 2016. p. 20–4.
- 114. Hughes PE, Caenepeel S, Wu LC. Targeted Therapy and Checkpoint Immunotherapy Combinations for the Treatment of Cancer [Internet]. Vol. 37, Trends in Immunology. Elsevier Ltd; 2016 [cited 2020 Apr 3]. p. 462–76. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27216414
- 115. Dempke WCM, Fenchel K, Uciechowski P, Dale SP. Second- and third-generation drugs for immuno-oncology treatment—The more the better? Vol. 74, European Journal of Cancer. Elsevier Ltd; 2017. p. 55–72.
- 116. Quesada AE, Assylbekova B, Jabcuga CE, Zhang R, Covinsky M, Rios A, et al. Expression of Sirt1 and FoxP3 in classical Hodgkin lymphoma and tumor infiltrating lymphocytes: Implications for immune dysregulation, prognosis and potential therapeutic targeting. Int J Clin Exp Pathol. 2015;8(10):13241–8.
- 117. Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, et al. Use of chemotherapy plus a monoclonal antibody against her2 for metastatic breast cancer that overexpresses HER2. N Engl J Med. 2001 Mar 15;344(11):783–92.
- 118. Dong H, Strome SE, Salomao DR, Tamura H, Hirano F, Flies DB, et al. Tumor-associated B7-H1 promotes T-cell apoptosis: A potential mechanism of immune evasion. Nat Med. 2002;8(8):793–800.
- 119. Powles T, Eder JP, Fine GD, Braiteh FS, Loriot Y, Cruz C, et al. MPDL3280A (anti-PD-L1) treatment leads to clinical activity in metastatic bladder cancer. Nature [Internet]. 2014 Nov 27 [cited 2020 Mar 25];515(7528):558–62. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25428503